Plant Physiology Preview. Published on June 17, 2015, as DOI:10.1104/pp.15.00731

Running head: Leaf anatomy and water transport

Corresponding author: Tom Buckley

email: *t.buckley@sydney.edu.au*
phone: +61 481 009 451
postal address: University of Sydney
12656 Newell Hwy
Narrabri, NSW 2390
Australia

How does leaf anatomy influence water transport outside the xylem?

Thomas N. Buckley [1]
Grace P. John [2]
Christine Scoffoni [2]
Lawren Sack [2]

[1] IA Watson Grains Research Centre, Faculty of Agriculture and Environment, The University of Sydney, 12656 Newell Hwy, Narrabri, NSW 2390 Australia

[2] Department of Ecology and Evolutionary Biology, University of California, Los Angeles, Los Angeles, California, USA 90095

Summary of most important findings:

Anatomical data from diverse species, applied to a novel integrative model, elucidates the mechanistic basis of differences in water transport outside the xylem in leaves.

Financial sources:

This work was supported by the US National Science Foundation (Award #1146514). TNB was also supported by the Australian Research Council (DP150103863 and LP130101183), the Bushfire and Natural Hazards Cooperative Research Centre and the Grains Research and Development Corporation.

Corresponding author: Tom Buckley
email: *t.buckley@sydney.edu.au*

Abstract

Leaves are arguably the most complex and important physico-biological systems in the ecosphere. Yet water transport outside the leaf xylem remains poorly understood, despite its impacts on stomatal function and photosynthesis. We applied anatomical measurements from 14 diverse species to a novel model of water flow in an areole (the smallest region bounded by minor veins) to predict the impact of anatomical variation across species on outside-xylem hydraulic conductance (K_{ox}). Several predictions verified previous correlational studies: e.g., (i) vein length per unit area is the strongest anatomical determinant of K_{ox}, due to effects on hydraulic pathlength and bundle sheath (BS) surface area; (ii) palisade mesophyll remains well hydrated in hypostomatous species, which may benefit photosynthesis, (iii) BS extensions (BSEs) enhance K_{ox}, and (iv) the upper and lower epidermis are hydraulically sequestered from one another despite their proximity. Our findings also provided novel insights: (v) the BS contributes a minority of outside-xylem resistance; (vi) vapour transport contributes up to two-thirds of K_{ox}; (vii) K_{ox} is strongly enhanced by proximity of veins to lower epidermis; and (viii) K_{ox} is strongly influenced by spongy mesophyll anatomy – decreasing with protoplast size and increasing with airspace fraction and cell wall thickness. Correlations between anatomy and K_{ox} across species sometimes diverged from predicted causal effects, demonstrating the need for integrative models to resolve causation. For example, (ix) K_{ox} was enhanced far more in heterobaric species than predicted by their having BSEs. Our approach provides detailed insights into the role of anatomical variation in leaf function.

Keywords: leaf anatomy, hydraulic efficiency, leaf traits, stomatal conductance, vascular transport

Introduction

Leaf hydraulic conductance (K_{leaf}) varies widely among species (Brodribb et al., 2005; Sack and Holbrook, 2006; Sack and Scoffoni, 2013). Because the resistances inside and outside the leaf xylem (R_x and R_{ox}) also vary widely, and are, on average across species, of a similar order of magnitude (Sack and Holbrook, 2006), both vein traits and mesophyll anatomy have potentially strong influences on K_{leaf}. This variation has important implications for the ecological consequences of leaf anatomy, for the coordination of water status and water flow across scales in plants and for stomatal regulation, which may be influenced by micro-scale variations in leaf water potential (Buckley 2005, Mott 2007). However, the mechanistic basis of variation in the hydraulic conductance outside the xylem (i.e., across the bundle sheath to the sites of evaporation), $K_{ox} = 1/R_{ox}$, is poorly understood (see Table I for a list of parameters and symbols used in this study).

A strong empirical correlate of K_{leaf} is vein length per unit leaf area (VLA) (Sack and Frole, 2006; Brodribb et al., 2007), which is predicted to increase both K_x and K_{ox} – the former, by providing additional parallel flow paths through the vein system, and the latter, by decreasing horizontal path length for water transport from the minor veins to the sites of evaporation. High VLA may also be associated with shorter vertical path length if VLA is negatively correlated with leaf thickness, as is observed within certain species sets and lineages but not others (Noblin et al., 2008; Sack et al., 2013; Sack et al., 2014; Zwieniecki and Boyce, 2014). However, K_{ox} might be correlated with VLA due to the influence of other traits that are structurally associated with veins and are positively correlated with K_{leaf}, such as the size and hydraulic permeability of bundle sheath (BS) cells and the presence and size of bundle sheath extensions (BSEs). Mesophyll tissue thickness and the ratio of spongy to palisade mesophyll tissue thickness are also both correlated with K_{leaf} (see Sack et al., 2015 for a comprehensive review of anatomical determinants of K_{leaf}). Additionally, across species, mesophyll anatomy, venation architecture, stomatal conductance and K_{leaf} tend to be inter-correlated (Sack et al., 2003; Aasamaa et al., 2005; Brodribb and Jordan, 2008; Carins Murphy et al., 2012; Brodribb et al., 2013; Feild and Brodribb, 2013; Carins Murphy et al., 2014). Thus, many of the key anatomical traits that may influence K_{ox} tend to be highly correlated across species (John et al., 2013), making it difficult to infer causal relationships.

Clarity on these issues requires application of detailed anatomical data to a model that links leaf anatomy to the physics of water transport, allowing testable predictions about K_{ox} to be generated from alternative hypotheses about water movement beyond the xylem. Earlier models demonstrated that leaf anatomy can play a critical role in determining the sites of evaporation and major resistances within the leaf and the consequences of these features for stomatal regulation (e.g., Meidner, 1976; Tyree and Yianoulis, 1980). More recent work has led to new insights, as well as new questions, about the nature and role of vapour phase water transport within the leaf, highlighting the need to better represent the anatomical structure of the mesophyll and surrounding air spaces in models (Rockwell et al., 2014; Buckley, 2015). The latter study made steps towards a more anatomically explicit model of leaf water flow, and presented a first analysis of the effects of epidermal and mesophyll anatomy on partitioning of flow among apoplastic, symplastic and gas phase transport modes. However, that analysis did not include several key tissues (the BS and BSEs), and it did not attempt to integrate across tissues, transport modes and directions of flow to estimate values of K_{ox} comparable to experimental data. A new approach was needed to refine and test hypotheses for the influence of anatomy on water flow outside the xylem.

The objective of this study was to test hypothesized relationships between leaf anatomy and outside-xylem water transport by extending the framework of Buckley (2015) to create a new, spatially explicit model of outside-xylem water transport, MOFLO (mesophyll and outside-xylem flow), that includes all leaf tissues, including BS and BSEs. MOFLO computes K_{ox} and its BS and outside-BS components (K_b and K_{ob}, respectively) by simulating steady-state water transport outside the xylem in an areole (the smallest region of a leaf bounded by minor veins). We estimated 34 anatomical parameters from light micrographs of transverse leaf sections from 14 species diverse in phylogeny, leaf structure and ecology, and assessed the mechanistic influence of these parameters on K_{ox} by varying each parameter in isolation in the model while holding the others constant. We performed a range of alternative simulations to address uncertainty in parameters that could not be confidently estimated by light microscopy. We used these simulations to address five interrelated questions: (1) Where are the major resistances located outside the xylem (i.e., in which tissues, and in which type of flow pathways), and

135 particularly, how much resistance is contributed by the BS? (2) How do BSEs affect K_{ox}? (3)
136 How do other cell and tissue anatomical traits influence K_{ox} and K_{leaf}? (4) Can these influences
137 explain previously described correlations of anatomical traits, and particularly VLA, with K_{leaf}?
138 (5) What are the roles of gas-phase transport, temperature and vertical temperature gradients in
139 determining K_{ox}?
140
141 **Results**
142 *Comparison of simulated values of K_{ox} across species with measured values*
143 Observed K_{ox} ranged from 3.5 to 54.3 mmol m^{-2} s^{-1} MPa^{-1} across the eight species for which
144 measurements were available (Table II, Figure 1). The mean and median simulated K_{ox} across
145 those eight species (16.8 and 13.6 mmol m^{-2} s^{-1} MPa^{-1}, respectively), were greater than, but of
146 similar order of magnitude to the mean and median observed K_{ox} (11.9 and 5.4 mmol m^{-2} s^{-1}
147 MPa^{-1}, respectively) (Table II, Figure 1). For seven of the eight species measured, the observed
148 values fell between the "low" and "high" simulated values from simulation set (i), which used a
149 wide span of values for each of the six "unknown" parameters of leaf design (Table III). The
150 exception was *Salvia canariensis*, for which measured K_{ox} exceeded the "high" simulated value.
151 The measured and modeled values of K_{ox} were uncorrelated across species ($p > 0.05$; not shown),
152 which was to be expected, given that our modeled estimates of K_{ox} are based on assumed values
153 for several parameters whose true values are unknown and may differ across species.
154
155 *Modeling the water potential drawdown outside the xylem*
156 Figure 2 shows an example of the simulated distribution of water potential drawdown outside the
157 xylem ($\delta\psi$) in a transverse section of a radially symmetrical areole, for one species,
158 *Comarostaphylos diversifolia*, using default values for all parameters (Tables III-IV). The
159 drawdown increases from the bundle sheath (at the left-hand edge of the figure, in rows 18-22),
160 to the lower (abaxial) epidermis at the center of the areole (the bottom right corner of the figure).
161 Although the drawdown exceeds –2.2 MPa, the volume-weighted average drawdown is only –
162 0.60 MPa (or –0.63 MPa excluding the bundle sheath itself). One reason for this difference is
163 that much of the leaf's water is in palisade mesophyll, which is outside of the main pathways for
164 water flow from the xylem to the transpiring epidermis and consequently experiences little

drawdown. In this example, simulated K_{ox} was 7.9 mmol m^{-2} s^{-1} MPa^{-1}, K_b was 19.1 mmol m^{-2} s^{-1} $MPa^{-1}$and K_{ob} was 13.4 mmol m^{-2} s^{-1} MPa^{-1}.

Partitioning hydraulic resistance outside the xylem

Across all 14 species, simulated K_{ox} ranged from 4.0 to 28.6 mmol m^{-2} s^{-1} MPa^{-1}, with a median of 9.0 and mean of 11.8 (Table V). Simulated K_{ob} varied from 4.8 to 47.4 (median 13.2) and K_b varied from 7.1 to 136 (median 41.6). On average, for default parameter values, most outside-xylem resistance occurred outside the bundle sheath: although the BS contribution ranged from 12 to 71%, the median was 18%.

The importance of tissue types and transport modes in outside-xylem water transport

Tissue types and transport modes varied widely in their contributions to outside-xylem water transport. On average across species, the bulk conductivity (k, flow per unit water potential gradient, per unit bulk tissue area; mol s^{-1} m^{-2} (MPa m^{-1})$^{-1}$) was greatest in the lower epidermis and lowest in the palisade mesophyll (for horizontal transport), followed closely by spongy mesophyll transport (Figure 3). Bulk conductivity in BSEs and across the BS itself were more than double that of the spongy mesophyll (Figure 3). Apoplastic pathways provided most transport in all tissues, although transmembrane and (isothermal) gas phase transport modes together contributed nearly half of the bulk conductivity in the spongy mesophyll. (The roles of anisothermal vertical gas phase transport driven by temperature gradients, and of temperature itself, are discussed further below.)

Effect of changes in six "unknown" parameters: apoplastic pore diameter, cell membrane permeability, BS suberization, palisade connectivity, cell wall thickness and vertical temperature gradient

Outside-xylem hydraulic conductance was highly sensitive to the values of parameters that could not be estimated confidently, which we refer to here as "unknown" parameters (listed in Table III). Poiseuille radius of apoplastic nanopores (R_a) (Figure 4). Under default values for other parameters, K_{ox} increased by 668% when R_a increased from 3 to 10 nm, and decreased by 71% when R_a decreased from 3 to 0 nm (Figure 4). However, K_{ox} was less sensitive to the osmotic water permeability of cell membranes (P_m) under default values for other parameters, increasing

only 52% when P_m was increased four-fold from 40 to 160 μm s^{-1}, and decreasing just 18% when P_m was reduced from 40 to 0 μm s^{-1} (Figure 4). However, if the BS apoplast was assumed to be suberized, then R_a and P_m had similar influences on K_{ox} (Figure 4).

The fraction of horizontal palisade surface area in contact with adjacent palisade cells (f_{cph}) had little effect on K_{ox}, which increased only 45% when f_{cph} increased from 0% and 100% of the apparent value measured by light microscopy (i.e., when ρ_{fcph} increased from 0 to 1); furthermore, most of this increase occurred below $\rho_{fcph} = 0.2$ (Figure 5). Cell wall thickness was far more important in determining K_{ox}: K_{ox} increased by 400% when cell wall thicknesses used in simulations were increased from 20% to 100% of the values determined by light microscopy (i.e., when ρ_{ta} was increased from 0.2 to 1.0) (Figure 5).

Mean K_{ox} across species was strongly enhanced by the presence of a vertical temperature gradient within the leaf: doubling the gradient from its default value of 0.1 ^{o}C increased K_{ox} by 75%, and eliminating the gradient reduced K_{ox} by 27% (Figure 6) (note that 0.1^{o}C was the average temperature drop from the point of maximum temperature to the lower epidermis across species; in practice, we used the same gradient [$4.6 \cdot 10^{-4}$ ^{o}C μm^{-1}] for all species, so that the absolute temperature drop varied across species in relation to leaf thickness). Comparing these simulations to another that excluded gas phase transport altogether, we calculated that the average gas phase contribution to K_{ox} increased from 16% to 65% as the temperature gradient increased from 0 to 0.2^{o}C.

We also assessed the effect of temperature itself, as distinct from temperature gradients. K_{ox}, K_b and K_{ob} all increased strongly with temperature (Figure 7), but the relative increases in K_{ox} and K_{ob} were far greater than that for K_b: K_{ox} and K_{ob} increased by 286% and 378%, respectively, as temperature increased from 10 to 40°C, whereas K_b only increased by 81% over the same temperature range (note that the effect of temperature on K_b results only from changes in the diffusivity of liquid water in water, D_{ww}, because our model did not include any gas phase water transport across the BS due to the lack of airspaces in the BS).

Changes in the six "unknown" parameters did not, in most cases, result in substantial changes in the partitioning of hydraulic resistance outside the xylem, which proved robust across most simulations: less than 25% of outside-xylem resistance was contributed by the BS under any tested combination of values for R_a and P_m, provided the BS apoplast was not assumed to be suberized (Figure 8). When a BS Casparian strip was included in the simulations (thus preventing apoplastic transport across the BS), the BS accounted for nearly 40% of total outside-xylem resistance under default values for other parameters, and up to 75% for high R_a (10 nm) and low P_m (20 μm s^{-1}) (Figure 8). However, changes in ρ_{ta} had little effect on the percentage of outside-xylem resistance in the BS, which decreased from 25.2% to 23.2% as ρ_{ta} increased from 0.2 to 1.0 (not shown).

Functional consequences of "known" anatomical traits on K_{ox}: VLA, vein positioning, leaf thickness, bundle sheath extensions and leaf airspace fraction

Table VI lists standardised slopes for linear regressions between each anatomical parameter and modeled K_{ox}. By far the strongest influence of leaf anatomy on K_{ox} was that of vein length per unit leaf area (VLA): K_{ox} increased 121% with a doubling of VLA (Figure 9), due in part to the effect of VLA on bundle sheath surface area per unit leaf area (which affects K_b, Figure 9), and in part to the fact that VLA reduces the horizontal pathlength for water transport to the transpiring epidermis (which affects K_{ob}, Figure 9). The pathlength effect was stronger than the BS area effect (increasing VLA increased the proportion of outside-xylem resistance in the BS; not shown). The VLA effect was over three times stronger than the next strongest anatomical effects: the increase in K_{ox} resulting from greater relative proximity of the vascular bundle to the abaxial epidermis (represented here as the ratio of distances between the BS and the upper vs lower epidermis; Figure 10), and the decrease in K_{ox} caused by increasing spongy mesophyll cell radius (Figure 11). (The spongy cell radius effect arises because of the dominance of apoplastic transport: if cell radius increases without a concomitant increase in wall thickness, the apoplastic fraction of available transport area declines.) For both of the latter effects, K_{ox} changed by approximately one-third with a doubling of the parameter value (Table VI).

Across species, K_{ox} was uncorrelated with leaf thickness, and leaf thickness had a smaller mechanistic influence on K_{ox} (doubling thickness reduced K_{ox} by 18%; Figure 10; Table VI) than

the relative proximity of the vascular bundle to the lower epidermis. The lack of a cross-species correlation between leaf thickness and modeled K_{ox} may partly reflect a positive correlation between leaf and cell wall thicknesses in our species (not shown), which would tend to counteract the effect on K_{ox} of increased vertical pathlength in thicker leaves.

Eight of our 14 species were heterobaric (they possessed BSEs), and six were homobaric. We assessed the mechanistic effect of BSEs on K_{ox} by comparing standard simulations with another set of simulations in which BSEs were replaced with mesophyll tissue in the model. These simulations found that BSEs directly increased K_{ox} by 10% on average across the eight heterobaric species (Figure 12). However, K_{ox} was 34% greater in heterobaric than homobaric species (Figure 12), which suggests the enhancement of K_{ox} in heterobaric species is mostly due to factors other than BSEs themselves.

Correlations of anatomy across species with K_{ox} – divergence from mechanistic relationships

In each of the cases described above, the correlation between each parameter and the simulated values of K_{ox} across species was in the same direction as the mechanistic effect. The opposite was true for several other parameters, however. For example, the mechanistic effect of the fraction of spongy mesophyll cell area in contact with adjacent cells (f_{cs}) was positive – simulated K_{ox} increased 24% with a doubling of f_{cs} – whereas the correlation across species was strongly negative (Figure 11, Table VI). The converse was true for the ratio of palisade to spongy mesophyll thickness: the mechanistic effect of this ratio on K_{ox} was weakly negative, but the correlation across species was strongly positive (Figure 11, Table VI). Spongy mesophyll airspace fraction (p_s) also had a positive mechanistic influence on K_{ox} (Figure 11), with K_{ox} increasing 35% as p_s increased from 0.1 to 0.6, whereas these variables were uncorrelated across species (Table VI).

Discussion

We elucidated and addressed key hypotheses for the anatomical basis of outside-xylem hydraulic conductance, K_{ox}, by applying measured variations in leaf anatomy across a set of very diverse species (Table VII) to a novel computational model, MOFLO. Our analysis led to several

predictions consistent with previous work, but equally, to a number of surprising novel predictions. We addressed several questions, discussed below.

Where are the major resistances outside the xylem?

Our simulations converged in showing that most resistance beyond the xylem occurs in the spongy mesophyll, and that the bundle sheath (BS) contributes a minority of outside-xylem resistance. The spongy mesophyll is intrinsically more resistive than other tissues because its airspace fraction is high (averaging 37% across species, nearly twice that of the palisade) and its cell-to-cell connectivity is low (an average of 26% of spongy cell surface is in contact with other cells), both of which reduce the area effectively available for liquid-phase flow. Our calculations suggest the epidermis is over three times as conductive on a bulk area basis than spongy mesophyll, on average across our 14 study species. Only horizontal transport in the palisade has a lower bulk conductivity than the spongy mesophyll, but this has little impact on K_{ox} because most water flows through the spongy mesophyll in hypostomatous species (12 of the 14 species in this study).

The true contribution of the BS to outside-xylem resistance remains somewhat ambiguous due to uncertainty about the occurrence of a suberized layer ("Casparian strip") in BS cell walls. Such a strip would greatly reduce apoplastic conductivity across the BS, rendering the BS analogous to the root endodermis, and its presence is one of the major outstanding questions in leaf design. Previous studies have suggested a BS Casparian strip in certain grass species, plantagos and at least several other taxa (Lersten, 1997; Mertz and Brutnell, 2014), and the expression of similar genes during development in BS and root endodermis suggests functional similarities (Slewinski et al., 2012). In any case, even when the model was modified to include a BS Casparian strip, the average BS contribution to outside-xylem resistance only increased from 10% to 37% under default values for other parameters. Thus, we tentatively conclude that the BS contributes a significant but minority share of outside-xylem resistance.

Does liquid flow outside the xylem follow apoplastic and/or transmembrane routes?

Previous studies using staining or conceptual modeling have reached differing conclusions about the relative importance of transport across living cells or around them, in the apoplast.

Apoplastic tracer studies (Canny 1986) and discovery of aquaporins (Agre et al 1993, Chrispeels and Agre 1994) have promoted the view in recent years that transmembrane flow may dominate outside-xylem transport (Tyree et al 1981, 1999, Sack et al 2005), at least in the light, when aquaporins may be activated (Cochard et al., 2007). However, a theoretical study by Buckley (2015) that used membrane permeability values from published studies carried out on illuminated leaves concluded that apoplastic transport should dominate. MOFLO extends upon that study, and similarly predicted that that apoplastic bulk flow contributes the majority of K_{ox} (68% on average across species), thus dominating both transmembrane and gas phase pathways under most conditions. This is due to the intrinsically greater efficiency of apoplastic bulk flow than either liquid or gas phase diffusion. Although our LM-based measurements of cell wall thickness (which strongly determine apoplastic conductance) were much greater than most published estimates for other species, this does not explain the model's predictions concerning apoplastic transport, because by default we reduced our LM-based estimates of cell wall thicknesses by 80% before applying them to the model ($\rho_{ta} = 0.2$). Transmembrane pathways contributed only 19% of K_{ox} on average, and this fraction was smaller still (6%) if LM-based cell wall thicknesses were used. (The contribution of gas phase pathways is discussed below.)

These conclusions assume that bulk flow in the apoplast can be modeled using Poiseuille's Law, which is derived from the Navier-Stokes equations of continuum fluid mechanics. Continuum hydrodynamics is valid provided the flow channels are large relative to the chemical species. The relevant size measure for liquid water molecules in this context is the lattice spacing, which is approximately 0.31 nm. Eijkel & Van Den Berg (2005) note that "friction is seen to increase from the macroscopic [continuum-derived] value when the separation between two surfaces becomes less than, roughly, ten molecular layers", or ~3 nm in this case. This is identical to the low end of the range estimated by Buckley (2015) for the diameter of channels for water flow created by spaces between adjacent microfibrils or bundles of microfibrils in the apoplast (3–20 nm) based on published measurements of cell wall microstructure (McCann et al., 1990; Fleischer et al., 1999; Fahlén and Salmén, 2004; Kennedy et al., 2007), which suggests the continuum approximation is probably reasonable for apoplastic transport.

The framework developed by Buckley (2015) included a term for diffusive resistance across the cellular interior ("transcellular resistance") in series with transmembrane resistance. Further thought and discussions with colleagues led us to conclude that any water transport across the cellular interior probably occurs mostly by bulk flow, provided the flow area consists of channels much greater than the 0.31 nm lattice spacing of water. Even if those channels had a typical radius similar to those in the adjacent cell walls, transcellular resistance would be on the same order of magnitude as apoplastic resistance (and thus far smaller than transmembrane resistance) if the transcellular area available for water flow were similar to the apoplastic flow area. Regardless, if this is incorrect and transcellular resistance is large, that would only strengthen our conclusion that apoplastic transport dominates outside-xylem water transport.

The effect of bundle sheath extensions

Previous studies that inferred the effect of BSEs on K_{leaf} from anatomy, simpler hydraulic models, K_{leaf} responses to light and stomatal responses to evaporative demand in hetero- vs homobaric species have hypothesized that BSEs are a major route for water flow from the veins to the epidermis and thence to the stomata (Wylie, 1952; Scoffoni et al., 2008; Buckley et al., 2011; Sommerville et al., 2012; Zsögön et al., 2015). MOFLO allowed us to directly quantify the effect of BSEs on K_{ox} by replacing BSEs with mesophyll tissue in the model. The results suggested BSEs enhance K_{ox} by an average of 10% across the eight heterobaric species in this study. However, simulated K_{ox} was 34% greater in these species than in the six homobaric species. This finding suggested that the presence of BSEs is correlated with one or more other traits that also enhance K_{ox}. The only anatomical parameter that differed significantly between heterobaric and homobaric species in our dataset was spongy mesophyll cell radius, r_s ($p < 0.05$, 2-tailed t-test with unequal variances): r_s was greater in homobaric species (21 ± 3 vs 12 ± 2 μm). This is consistent with our mechanistic trait analysis, which predicted that K_{ox} should decrease by 30% for a doubling of r_s (Table VI).

Effects of cellular dimensions on K_{ox}

Most individual anatomical traits affected K_{ox} only weakly. The major exceptions involved spongy mesophyll anatomy, which had much larger influences than palisade anatomy because most of our study species (12 of 14) were hypostomatous, so little water transport occurs through

379 the upper half of the leaf. The apparent effect of spongy mesophyll radius, r_s, in our trait analysis
380 arose because when all other parameters are held constant, increasing r_s increases the
381 transmembrane fraction of the total cross-sectional area available for flow, which decreases the
382 apoplastic fraction, in turn decreasing K_{ox}. However, r_s is often correlated with spongy mesophyll
383 cell wall thickness across species (e.g., John et al 2013), which would tend to reduce the direct
384 effect of r_s. Another explanation for the similarity between the correlative and mechanistic
385 relationships that we found between r_s and K_{ox} (Fig 10b) is that r_s was negatively correlated with
386 VLA and with the relative proximity of vascular bundles to the lower epidermis ($r^2 = 0.25$ and
387 0.61, respectively; $p < 0.0001$ for both), both of which had positive mechanistic effects on K_{ox}, as
388 discussed below. A similar negative correlation between VLA and the sizes of mesophyll and
389 epidermal cells was previously reported to hold across species of Proteaceae by Brodribb *et al*.
390 (2013).
391
392 *Effects of VLA, leaf thickness and distance from vascular bundles to epidermis*
393 The specific role of VLA in increasing outside-xylem flow has been a topic for debate. Sack &
394 Frole (2006) suggested that higher VLA led to shorter horizontal flow distances, increasing K_{leaf}.
395 This was also found by Brodribb et al. (2007), who additionally hypothesised that a shorter
396 vertical distance between vein and epidermis would also increase K_{leaf}. Indeed, because high
397 VLA leaves are often thinner as well – a correlation that has been hypothesized to be optimal for
398 water transport based on modeling using artificial leaf assemblies (Noblin et al., 2008) – a high
399 VLA would also correspond to such shorter vertical distance. Brodribb et al. (2007) combined
400 the hypothesized effects of horizontal and vertical distances in their variable D_m, representing a
401 diagonal distance from veins to epidermal evaporating sites, and reported a strong correlation
402 between D_m and leaf hydraulic resistance, which was mostly driven by VLA. However, Sack et
403 al. (2013) suggested that greater leaf thickness should contribute to higher K_{ox} given the greater
404 number of parallel pathways for horizontal transport to the sites of evaporation, provided those
405 sites are distributed throughout the leaf. MOFLO allowed us to test these putative mechanisms.
406 We found that increasing VLA, reducing total leaf thickness and reducing the relative distance of
407 vascular bundles from the lower epidermis all increased K_{ox} in the model because of their effects
408 on reducing flow pathlengths, although the effect of VLA was by far the strongest and that of
409 leaf thickness the weakest of the three. The model found that VLA affects K_{ox} in two ways: by

increasing BS surface area per unit leaf area, which affects K_b, and by decreasing horizontal pathlength, which affects K_{ob}. Although both effects were quite strong, the pathlength effect was stronger (K_{ob} and K_b increased 113% and 94%, respectively, with a doubling of VLA; Figure 9).

The model also found a negative mechanistic effect of vertical pathlength (as influenced by either total leaf thickness or relative vein-to-epidermis distance), but these effects were only one-sixth and one-third as strong, respectively, as the horizontal-distance effect of VLA (Table VI). The main reason for the smaller effect of changes in vertical pathlength (i.e., of leaf thickness) than horizontal pathlength (i.e., VLA) on K_{ox} is that adding vertical layers simultaneously also reduces the horizontal resistance by providing additional parallel pathways for horizontal transport (Sack et al. 2013). In contrast to its mechanistic effect, we found that leaf thickness was not significantly correlated with simulated K_{ox} across our species, due to compensating effects of other parameters that covaried with leaf thickness. For example, leaf thickness was strongly and positively correlated with cell wall thickness in each tissue type (r^2 between 0.33 and 0.69, $p <$ 0.0001 in all cases; not shown), all of which had strongly positive mechanistic effects on K_{ox} (Table VI). These results verify that the often-observed correlation between K_{leaf} and VLA is mechanistic in origin (Sack and Frole, 2006; Brodribb et al., 2007; Brodribb and Jordan, 2008; Carins Murphy et al., 2012; Feild and Brodribb, 2013; Carins Murphy et al., 2014), and they further suggest that the horizontal pathlength component of the VLA effect is more important than the vertical component.

The role of gas phase transport and vertical temperature gradients

Recent work has raised the possibility that gas phase water transport contributes a substantial fraction of the total conductance for water movement through the mesophyll – perhaps comparable in magnitude to that provided by liquid phase pathways – particularly for vertical transport in the presence of large vertical temperature gradients (Rockwell et al., 2014; Buckley, 2015). Our analysis extended that work by providing, for the first time, an integrated measure of K_{ox} that includes both horizontal and vertical components of gas phase transport, all in the same leaf area-based hydraulic conductance units. The model found that gas phase transport contributed an average of 39% of K_{ox} across species under default conditions (which include a baseline temperature of 25°C and a vertical temperature gradient of 0.1^{o}C). This rose to 65% for

a gradient of 0.2°C and fell to 16% for zero gradient. Thus, we conclude that the contribution of vapour transport within the leaf to the apparent conductance for water transport can be quite substantial.

This has several implications for interpreting leaf function and gas exchange. First, it implies that generation of vertical temperature gradients by preferential absorption of light near the upper leaf surface can enhance K_{ox} greatly – by over 20% for 0.1°C gradients or 40% for 0.2°C gradients. This corresponds to average 16% and 31% enhancements of K_{leaf}, respectively, across the eight species in our dataset for which we measured K_{ox}. These effects could contribute to observed effects of light on K_{leaf}, in addition to other mechanisms such as increased aquaporin activity (Cochard et al., 2007; Scoffoni et al., 2008; Voicu et al., 2009).

Second, a major role for vapour transport implies that a great deal of water may evaporate from cells deep within the leaf. This contrasts with some earlier conclusions (e.g., Tyree and Yianoulis, 1980) that the great majority of evaporation occurs from cells very close to the stomatal pore, but it is consistent with conclusions of Boyer (1985) based on measurements of vapour diffusion pathlength by Farquhar and Raschke (1978). The question of where evaporation occurs within the leaf has remained one of the most challenging and critically important in plant water transport for decades (Meidner, 1983; Barbour and Farquhar, 2004), and demands further discussion here. In the context of water transport, evaporation represents a shift of water from a liquid pathway to a gas phase pathway. Water flow will distribute itself across pathways so as to minimise total resistance; therefore, some water will switch from a liquid to a gas phase pathway whenever the gas phase conductance increases relative to the liquid phase conductance (Buckley, 2015). Thus, *evaporation should occur wherever the gas-phase fraction of total conductance increases along a trajectory of flow* (a pathway normal to isoclines of water potential). . That fraction increases substantially in three areas: (1) at the outer margin of the bundle sheath (where the fraction rises from zero to some positive value when water first encounters airspaces in the leaf), (2) at the boundary between palisade and spongy mesophyll (where the gas phase fraction increases due to increasing tissue airspace fraction and decreasing vertical liquid-phase conductance), and (3) at open stomatal pores, where the gas phase fraction approaches 100% (because all water exits the leaf as vapour). This suggests that evaporation is clustered in three

locations in hypostomatous leaves: the BS, the upper spongy mesophyll and surfaces immediately adjacent to open stomata. A similar argument would apply to amphistomatous species with spongy mesophyll in the center of the leaf, except that the prevailing direction of water flow would be from spongy into palisade mesophyll, implying that condensation rather than evaporation would occur at the spongy/palisade transitions. The liquid phase share of transport from those regions to the transpiring epidermes would thus be greater in amphistomatous species than in hypostomatous species (due to the greater liquid conductivity and smaller porosity of palisade as compared to spongy mesophyll), which in turn implies that a greater share of evaporation would occur from surfaces very close to the stomata in amphistomatous species.

The role of temperature itself

The direct effect of temperature on K_{ox} (independent of temperature gradients) was also substantial in the model: under otherwise default parameter values, K_{ox} increased 25% as leaf temperature increased from 25 to 30°C, and 233% for an increase from 25 to 40°C. This effect arises partly from the temperature dependence of liquid-phase conductivities (chiefly due to decreasing dynamic viscosity), but more so from increasing gas-phase conductivities (due to strong increases in both the molecular diffusivity of water vapour in air and the saturation vapour pressure). These direct temperature effects could further contribute to light responses of K_{leaf} in nature, where temperature usually increases with absorption of sunlight. A direct increase in K_{ox} with temperature could also help to sustain turgor when water loss increases as a result of leaf warming rather than drying of the air; such an effect may also help to explain positive correlations reported between K_{leaf} and transpiration rate (Simonin et al., 2014) in cases where changes in transpiration are temperature-driven.

Implications for stomatal sensing of leaf water status

Our model suggested that large water potential gradients could occur between the xylem and the most distal epidermal tissues: in the example shown in Figure 2 (for *Comarostaphylos diversifolia*), the drawdown from the xylem to the lower epidermis at the center of the areole was 3.7 times greater than the average drawdown outside the xylem. This ratio varied across species, reaching 6.3 in *Magnolia grandiflora*, and it was substantial at 2.2 even in the amphistomatous

species *Helianthus annuus*. These results support the hypothesis that a transpiring epidermis (and the stomatal guard cells embedded therein) may experience far greater swings in water potential in response to changes in transpiration rate than one would infer from changes in bulk leaf water potential. This may help to reconcile "isohydric" behaviour (near-homeostasis in ψ_{leaf}) with a mechanism for stomatal responses based on a feedback response to changes in water potential somewhere in the leaf (Sperry, 2000; Buckley, 2005). The large drawdowns predicted by the model also suggest that the upper and lower epidermes are in effect hydraulically sequestered from one another, which may help to explain the observation that stomata at one surface appear only minimally responsive to changes in transpiration rate at the other surface (Mott, 2007). We tested this idea directly in MOFLO by tripling transpiration rate at the upper surface of a simulated *H. annuus* leaf while holding transpiration constant at the other surface; the resulting change in water potential at the center of the areole in the upper epidermis was 4.3 times greater than in the lower epidermis (Figure 13).

Conclusions

Our novel analyses provide, for the first time, quantitative integration of the effects of leaf anatomy on water flow outside the xylem, in terms directly comparable to experimental data. Our model confirmed some earlier predictions about the relation of K_{ox} to leaf anatomy – including that VLA is the strongest anatomical determinant of K_{ox} and that BSEs and thermally-driven vapour transport through spongy mesophyll can enhance K_{ox} – but also provided novel insights, including that the BS probably contributes a minority of outside-xylem resistance, that higher K_{ox} in heterobaric species is mostly due to parameters other than BSEs, that vapour transport may constitute a majority of K_{ox} when large vertical temperature gradients exist in the leaf and that many cross-species correlations between K_{ox} and leaf traits are not mechanistic in origin. Our model provides strong insights into the coordinated function of the living leaf, a tool to explore the implications of variation in leaf anatomy and a baseline for future trait analyses.

Materials and Methods

Empirical measurements of outside-xylem hydraulic conductance (K_{ox})

We determined K_{ox} from measured whole-leaf and leaf xylem hydraulic conductance (K_{leaf} and K_x, respectively) for eight of our 14 study species (Table II). K_{leaf} was obtained from whole leaf

hydraulic vulnerability curves previously published using the evaporative flux method (Scoffoni et al., 2012; Scoffoni et al., 2015). Because K_{leaf} declines as water potentials become more negative, we calculated for each species the average K_{leaf} for the interval of leaf water potential near full hydration (we used 0 to -0.3, 0 to -0.5 or 0 to -1.0 MPa, depending on species, to capture the interval before strong decline in K_{leaf}: n = 5-12). K_x was obtained as previously described (Scoffoni et al., 2015) using the vacuum pump method. Briefly, minor veins of fully hydrated leaves were cut under water over a light bench to ensure no major veins were severed. Cuts were made in between about 95% of tertiary veins, yielding 5 to 33 cuts mm^{-2} depending on sample size (larger leaves have their major veins spaced further apart, so that fewer but longer cuts were made) (Sack et al., 2012; Scoffoni and Sack, 2015). These cuts were enough for water to move directly out of minor veins, and not through outside-xylem pathways (Sack et al., 2004; Nardini et al., 2005). After minor veins were cut, leaves were connected by tubing to a water source on a balance, and placed in a vacuum chamber. A steady flow rate was determined for five levels of partial vacuum (0.06, 0.05, 0.04, 0.03, and 0.02 MPa). K_x was calculated as the slope of the flow rate against pressure, corrected for leaf temperature, normalized for leaf area and averaged (n = 5-11). Outside-xylem hydraulic conductance (K_{ox}) was calculated using Eqn 1 and standard errors were obtained from propagation of error:

(1) $$K_{ox} = \left(K_{leaf}^{-1} - K_x^{-1}\right)^{-1}.$$

We note that estimates of K_{ox} thus depend on the accuracy of K_{leaf} values. In particular, the evaporative flux method requires steady state transpiration and stable leaf water potential to enable determination of K_{leaf}. We followed the procedure tested and established for a wide range of species in previous work (Scoffoni et al., 2008; Pasquet-Kok et al., 2010; Guyot et al., 2012; Scoffoni et al., 2012). In measuring K_{leaf}, thirty minutes was chosen as a minimum to ensure that leaves had acclimated to high irradiance and stomatal conductance had stabilized. Previous studies found these criteria to be sufficient for stabilization of E, water potential and K_{leaf}. Tests for any change in E, leaf water potential and K_{leaf} with measurement time (after stable flow was established) across leaves of a given species for seven species with a wide range of leaf capacitance showed no relationship of K_{leaf} to measurement time (Scoffoni et al., 2008; Pasquet-Kok et al., 2010).

Measurement of leaf anatomical traits

We used measurements of 34 leaf anatomical traits (Table IV) across 14 species as described by John et al. (2013), based on from light micrographs of fully hydrated leaves fixed in formalin-acetic-acid, embedded in LR white, cut in transverse 1 μm sections using glass knives in a microtome and imaged using a 20x or 40x objective.

Outline of the modeling approach

We created a model that uses anatomical measurements to calculate the hydraulic conductances of pathways outside the xylem in leaves. The model is adapted from the framework developed by Buckley (2015), which calculates horizontal and vertical components of hydraulic conductance in each of three tissue types distal to the bundle sheath (epidermis, palisade mesophyll and spongy mesophyll), and in each of three transport modes (apoplastic, transmembrane and gas phase). We extended the framework to include the bundle sheath itself and the bundle sheath extensions, applied it to a spatially explicit grid representing a single areole to compute the distribution of water potential across the areole, and used that distribution to compute total outside-xylem hydraulic conductance (K_{ox}) and its bundle sheath (K_b) and outside-bundle-sheath (K_{ob}) components.

The original framework of Buckley (2015) included a term for hydraulic resistance due to diffusion across the interior of each cell in series with the transmembrane resistance. Discussions with colleagues led us to recognise that water movement across the cellular interior may occur by bulk flow rather than by diffusion, and that the resulting transcellular bulk flow resistance would be negligible relative to the transmembrane resistance. We thus omitted the transcellular resistance from MOFLO. This is discussed further in the Discussion. We also assumed that the quantitative contribution of plasmodesmatal flow to transpired water movement is negligible, consistent with its narrow circular slit (of width 1-2 nm) available for water flow between the membrane at its perimeter and the interior desmotubule of the endoplasmic reticulum (Doelger et al., 2014).

The areole grid

We simulated a transverse section through a circular areole (the smallest region of a leaf bounded by minor veins) as a grid. Our results therefore apply to regions of the leaf bounded only by minor veins, and not by the lower-order (major) veins; although our model does not directly account for free-ending veinlets, the values of vein length per unit leaf area (VLA) used to estimate areole dimensions did include veinlets. This grid had 744 nodes: 24 horizontal (parallel to the epidermis) and 31 vertical (Figure 14). The aspect ratio of 24/31 was based on the average ratio of areole radius to leaf thickness across species (0.77 ± 0.07; mean ± SE). Each node represents a band of tissue delimited by outer and inner radii (horizontal distances from the areole center) and upper and lower depths (vertical distances from the upper leaf surface) (Figure 14). Representing circular bands of tissue as single nodes is equivalent to assuming that the areole is radially symmetrical. Areole radius was computed from VLA following previous models that considered the vein system as a square grid with unit edge length x; this implies each areole is uniquely associated with a vein length of $2x$ and an area of x^2, so VLA = $2x/x^2 = 2/x$ (Cochard et al., 2004; Sack et al., 2004). Equating this area with that of a circle of radius r_{areole} ($\pi \cdot r^2_{areole} = x^2$) gives $r_{areole} = x/\pi^{0.5} = 2/(\text{VLA}\cdot\pi^{0.5})$.

Each tissue band (node) in the grid was identified with a tissue type (BS, upper or lower BSEs, upper or lower epidermis, or palisade or spongy mesophyll). All bands in the top and bottom rows of the grid were identified as upper and lower epidermis, respectively, and all bands in the left-most column (which corresponds to the outer margin of the areole, aligned with the nearest minor vein) were identified as BS or either BSEs (in heterobaric species) or mesophyll (in homobaric species). All other tissue bands were identified as either spongy or palisade mesophyll based on measured anatomical proportions (Figure 14). Formulas for tissue identity at each band are given in the Supplemental Material.

The heights of the upper-and lower-most rows were taken as the measured thicknesses of the upper and lower epidermis, respectively; the height of each of the remaining 29 rows was set as 1/29 of the remaining leaf thickness. For homobaric species, which lack BSEs, all column widths were set at 1/24 of areole radius. For heterobaric species, which possess BSEs, the width of the outermost (left-hand) column was set equal to one-half of the measured BSE width (the other

half of BSE width would be associated with the next areole to the left), and the widths of all other columns were set at 1/23 of the remainder of areole radius. The resulting differences in tissue band dimensions among columns and rows were taken into account when computing the cross-sectional areas and flow pathlengths for connections between adjacent nodes; calculations involving BS nodes were further modified to account for the mapping of the elliptical cross-section of the BS onto a rectangular column of nodes (see Supplemental Material for more details).

Computing flows and water potentials in the grid

We computed the steady-state distribution of water potential across the grid on the basis of mass conservation. For each node *i*, an expression for mass balance can be written as a linear function of the water potentials of all nodes, in which the coefficients are hydraulic conductances between adjacent nodes. For example, the sum of all flows into node *i* from adjacent nodes must equal the net flow out of node *i* through stomatal transpiration:

(2) $$\sum (\psi_j - \psi_i) K_{ji} = E_i \quad ,$$

where ψ_j is the water potential at node *j*, K_{ji} is the conductance (mol s^{-1} MPa^{-1}) between nodes *i* and *j*, E_i is any loss of water from node *i* by stomatal transpiration (mol s^{-1}), and the sums are taken over all nodes in the grid (for nodes that are not directly connected to node *i*, the conductance K_{ji} will be zero). Water enters the grid from the xylem, which is treated as a "reference node" with a water potential of zero. This reference node is not part of the grid, but its existence and location are implicitly incorporated by including a term for xylem-to-bundle sheath hydraulic conductance (K_{xb}) in the equation for each bundle sheath node:

(3) $$\sum (\psi_j - \psi_b) K_{jb} + (\psi_x - \psi_b) K_{xb} = E_b \quad ,$$

where the subscript *b* denotes a bundle sheath node. In the presence of vertical temperature gradients within the mesophyll, the conductances for vertical connections between mesophyll nodes will include both an "anisothermal" gas phase component ($K_{aniso,ji}$), which depends on the

temperature difference between the two nodes, and an "isothermal" component ($K_{iso,ji}$) that does not. Rewriting Eqn 2 to separate these components gives

(4) $$\sum(\psi_j - \psi_i)K_{iso,ji} + \sum(\psi_j - \psi_i)K_{aniso,ji} = E_i .$$

$K_{aniso,ji}$ is given by Eqn 5 (which is based on Eqn 15 in Buckley 2015):

(5) $$K_{aniso,ji} = \frac{D_{wa}}{(\psi_j - \psi_i)R_{gas}}\left(\frac{p_{sat,j}}{T_j} - \frac{p_{sat,i}}{T_i}\right)\left(1 + \frac{v_w \psi_i}{R_{gas}T_j}\right)\left(\frac{\gamma_{ji}a_{ji}}{\beta_{ji}l_{ji}}\right),$$

where D_{wa} is the molecular diffusivity of water vapour in air, v_w is the molar volume of liquid water, p_{sat} and T are the saturation vapour pressure and absolute temperature, respectively (at nodes i or j as indicated by subscripts), R_{gas} is the gas constant, a_{ji} and l_{ji} are the area and pathlength for the connection between nodes j and i, and γ_{ji} and β_{ji} are unitless corrections that convert simple areas and pathlengths, respectively, to those actually experienced by moving water (see *Calculating the conductance matrix* below for details). The quantity $v_w \cdot \psi_i / R_{gas}T_j$ on the right-hand side is << 1 for typical leaf water potentials (e.g., this quantity is 0.0145 for ψ_i = -2 MPa and T = 298 K), so it can be omitted with negligible error. Omitting that quantity from Eqn 5 and multiplying both sides by ψ_j - ψ_i gives the anisothermal component of the vapour *flow* (mol s^{-1}) from node j to node i, $F_{aniso,ji}$, as

(6) $$F_{aniso,ji} \equiv (\psi_j - \psi_i)K_{aniso,ji} = \frac{D_{wa}}{R_{gas}}\left(\frac{p_{sat,j}}{T_j} - \frac{p_{sat,i}}{T_i}\right)\left(\frac{\gamma_{ji}a_{ji}}{\beta_{ji}l_{ji}}\right),$$

Note that $F_{aniso,ji}$ is identical to the term inside the second sum on the left-hand side of Eqn 4. Because $F_{aniso,ji}$ does not directly depend on water potentials, it can be moved to the right-hand side and combined with the stomatal transpiration flux to give a single term on the right-hand side of each linear equation:

682 (7) $$\sum(\psi_j - \psi_i)K_{iso,ji} = E_i - \sum F_{aniso,ji} \equiv e_i .$$

683

684 Equation 7 represents a system of linear equations that can be expressed more compactly in
685 matrix form, as the product of a square matrix of conductance coefficients (**K**) whose elements
686 are the $K_{iso,ji}$, and a scalar vector ($\boldsymbol{\delta\psi}$), whose elements are the water potentials at each node,
687 expressed relative to xylem water potential (i.e., the steady-state water potential drawdowns from
688 the xylem to each node), with a vector **e** comprising the e_i on the right hand side:

689

690 (8) $$\mathbf{K}(\boldsymbol{\delta\psi}) = \mathbf{e} .$$

691

692 This system can be solved for $\boldsymbol{\delta\psi}$ by multiplying the inverse of **K** by the vector **e**:

693

694 (9) $$\boldsymbol{\delta\psi} = \mathbf{K}^{-1}\mathbf{e} .$$

695

696 We generated the vector of transpiration rates (the components E_i of the vector **e**) by multiplying
697 a fixed and arbitrary transpiration rate per unit leaf area (E_{leaf} = 0.005 mol m^{-2} s^{-1}) by the
698 projected leaf area corresponding to each node at each transpiring leaf surface. For
699 amphistomatous species (*Helianthus annuus* and *Romneya coulterii*), we partitioned total
700 transpiration rate between the upper and lower leaf surfaces using the ratio of maximum stomatal
701 conductances at each surface (estimated as the ratio of the products of mean stomatal density and
702 mean inner pore length for each surface). We measured stomatal density by counting stomata in
703 each of three 400× fields of view in three leaves per surface, per species, and measured pore
704 lengths for four stomata in each field of view using ImageJ software. We thus estimated that
705 58.2% and 43.6% of transpiration occurred from the lower surfaces of *H. annuus* and *R. coulteri*,
706 respectively. All other species were hypostomatous, so we assumed all transpiration occurred
707 from the lower surface.

708

709 *Calculating the conductance matrix*

710 We generated the conductance matrix (**K**) as follows. First, we computed a set of *intrinsic*
711 *hydraulic conductivities*, κ (molar flow rates [mol s^{-1}] per unit water potential gradient [MPa m^{-1}]

per unit area [m^2]) for each transport mode (apoplastic, transmembrane and gas phase). The spatial dimensions in these conductivities represent the actual pathlengths and actual flow areas experienced by water moving in a particular tissue. Those pathlengths and areas often differ from the simple or "bulk" values that one would infer from bulk tissue geometry (for example, the apoplastic pathlength around a cylindrical cell is longer than the simple distance across that cell, and the area available for gas phase flow is smaller than the total cross-sectional area). The second step was therefore to compute correction factors for pathlength and area in each tissue type and flow direction. The area correction was the ratio of actual flow area to simple (bulk) flow area (γ) and the pathlength correction was the ratio of actual flow pathlength to simple (direct) flow pathlength (β). Third, for each transport mode in a given tissue and flow direction, we multiplied κ by γ and divided it by β to give the corresponding *bulk conductivity*, k:

(10) $$k = \kappa \cdot (\gamma / \beta)$$

Fourth, we summed these bulk conductivities across transport modes for each tissue type and flow direction. Finally, for each connection between a pair of nodes (j and i), we converted the appropriate total bulk conductivity to a conductance (K_{ji}, flow per unit water potential difference; mol s^{-1} MPa^{-1}) by multiplying it by the bulk flow area (a_{ji}) and dividing it by the direct flow pathlength (l_{ji}) appropriate to the connection between those nodes:

(11) $$K_{ji} = k \cdot (a_{ji} / l_{ji})$$

For connections between different tissue types (with bulk conductivities k_1 and k_2, say), we computed the total conductivity as $(0.5/k_1 + 0.5/k_2)^{-1}$. The K_{ji} comprise the elements of the conductance matrix **K** (denoted as $K_{iso,ji}$ in Eqn 7). We derive expressions for intrinsic hydraulic conductivities (κ) in the following section. Expressions for γ, β, a and l are derived in the Supplemental Material.

Calculating intrinsic hydraulic conductivities

We derived intrinsic conductivities from expressions given by Buckley (2015). Note that the term "conductivity" in that paper referred to flow per unit area, per unit water potential *difference* (mol s^{-1} m^{-2} MPa^{-1}), whereas in this paper, we use the term "conductivity" to describe a flow per unit area, per unit water potential *gradient* (mol s^{-1} m^{-2} [MPa m^{-1}]$^{-1}$ = mol s^{-1} m^{-1} MPa^{-1}). Thus, in this paper, conductances are computed by multiplying conductivities by flow areas and dividing them by flow pathlengths, as described earlier.

For diffusion across a single membrane, the flow per unit area per unit water potential difference is P_m/RT (cf. Eqn 1 in Buckley, 2015), where P_m is the osmotic water permeability of the membrane (m s^{-1}), R_{gas} is the gas constant (J mol^{-1} K^{-1} = Pa m^3 mol^{-1} K^{-1}) and T is the absolute temperature (K). To convert this to an intrinsic conductivity, it must be multiplied by one-half of the transcellular pathlength, L_c (m) (because two membranes are encountered for every bulk distance L_c travelled; the value of L_c differs among tissue types and flow directions). Thus, the intrinsic conductivity for transmembrane pathways is

$$\kappa_{mem} = \frac{L_c P_m}{2 R_{gas} T}. \quad (12)$$

The intrinsic conductivity for free diffusion of water, other than across membranes, is

$$\kappa_{diff} = \frac{D_{ww}}{R_{gas} T} \quad (13)$$

where D_{ww} is the molecular diffusivity for water in liquid water (m^2 s^{-1}). The intrinsic conductivity for bulk flow of water through cell walls with nanopores having an effective Poiseuille radius of R_a is

$$\kappa_{bulk} = \frac{R_a^2}{8 \eta v_w} \quad (14)$$

where η is the dynamic viscosity of water and v_w is the molar volume of liquid water. (Note that Buckley's (2015) analogous expression [his Eqn 8] also contains factors that appear in our area and pathlength correction factors [γ and β], which are derived in the Supplemental Material.) For gas phase transport (water vapour diffusion), the intrinsic conductivity (κ_{gas}) contains an "isothermal" term that does not depend explicitly on vertical temperature gradients in the leaf, and an "anisothermal" term that does depend on such gradients. The isothermal term is

$$\kappa_{gas,iso} = \frac{D_{wa} v_w p_{sat}}{\left(R_{gas} T\right)^2} \tag{15}$$

where D_{wa} is the molecular diffusivity of water vapour in air and p_{sat} is the saturation vapour pressure. The anisothermal term is

$$\kappa_{gas,aniso,ji} = \frac{D_{wa}}{\left(\psi_j - \psi_i\right) R_{gas}} \left(\frac{p_{sat,j}}{T_j} - \frac{p_{sat,i}}{T_i} \right) \left(1 + \frac{v_w \psi_i}{R_{gas} T_j} \right) \tag{16}$$

where the subscripts j and i refer to values at the nodes above and below the internodal connection for which $\kappa_{gas,aniso}$ is to be calculated. Equation 16 requires the vertical distribution of temperature to be specified. We assumed that temperature varied parabolically with depth in the leaf, relative to a maximum value of T_{max} at a relative depth of z_{max}, and such that the temperature drop from the maximum value to the lower surface was equal to an input parameter, ΔT. Thus,

$$T(z) = T_{max} - \Delta T \left(\frac{z - z_{max}}{1 - z_{max}} \right)^2 \tag{17}$$

where z is relative depth (z = 0 and 1 at the upper and lower leaf surfaces, respectively). For each species, we set ΔT proportional to leaf thickness, such that its default value was 0.1 ^{o}C for the mean leaf thickness of 292.5 µm. We assumed z_{max} = 0.25, based on Rockwell et al. (2014).

Computing integrated leaf-level hydraulic conductances

In experimental studies, K_{ox} is typically calculated from K_{leaf} and K_x using Eqn 1, and K_{leaf} in our study was determined using the evaporative flux method described above. We note that there are a number of other methods in use for determining K_{leaf}, such as the high pressure flow method (e.g., Yang and Tyree, 1994), the rehydration kinetics method (e.g., Brodribb and Holbrook, 2003), or the vacuum pump method (e.g., Martre et al., 2001) (see reviews of methods and their contrasting assumptions and difference in simulated flow pathways in Sack and Tyree, 2005; Flexas et al., 2013). Although several studies have shown that the different methods tend to yield similar maximum K_{leaf} values (Sack et al., 2002; Scoffoni et al., 2008), we highly recommend the use of the evaporative flux method for the most accurate representation of outside-xylem hydraulic pathways, since water movement in this method would most closely resemble that of a naturally transpiring leaf. In the evaporative flux method, K_{leaf} is defined as the ratio of E_{leaf} to the difference between stem ψ and bulk leaf ψ (ψ_{leaf}) of a leaf bagged during transpiration and then equilibrated. Generally, the equilibrated ψ_{leaf} is assumed to represent the volume-weighted average over the mesophyll cells in the transpiring leaf. This assumes that negligible water is taken up from the xylem to the mesophyll during equilibration, which would be the case if the open conduits in the petiole of the excised leaf contained negligible volume – an assumption that requires testing, given that many species have open vessels of several cm extending from the petiole into higher vein orders (Tyree and Cochard, 2003; Chatelet et al., 2011; Scoffoni and Sack, 2014). Even accepting this typical assumption, additional ambiguity in the partitioning of K_{ox} into BS and outside-BS components (K_b and K_{ob}, respectively) arises when K_{ox} is calculated using the bulk water potential of the entire symplast. As we show in the Supplemental Material, this can lead to spurious differences in K_{ob} between leaves even when those leaves have identical flow properties outside the BS. These artefacts can be traced to the fact that the bulk water potential used to compute K_{ox} includes some tissues that are proximal to the transport pathways that K_{ob} is meant to represent.

To allow simulated values of K_{ox}, K_b and K_{ob} to be interpreted as independent measures of outside-xylem, across-BS and outside-BS hydraulic conductances, respectively, we therefore defined these conductances, for modeling purposes, in terms of a water potential gradient whose endpoint is distal to the BS. Specifically, for modeling purposes we defined K_{ox} as

826 (18) $K_{ox} = E_{leaf} \big/ \left|\delta\psi_{ob}\right|$,

827

828 where $\delta\psi_{ob}$ is the volume-weighted average water potential drawdown from the end of the xylem
829 to all tissues distal to the bundle sheath, given by Eqn 19:

830

831 (19) $\delta\psi_{ob} = \sum_i v_i \cdot \delta\psi_i \Big/ \sum_i v_i$

832

833 where i is an index representing all non-BS nodes in the grid, v_i is the volume of liquid water in
834 the tissue band represented by node i, and $\delta\psi_i$ is the component of $\boldsymbol{\delta\psi}$ for node i (calculation of v_i
835 for each node in the grid is described in the Supplemental Material). We defined K_b as

836

837 (20) $K_b = E_{leaf} \big/ \left|\delta\psi_{bn}\right|$,

838

839 where $\delta\psi_{bn}$ is the volume-weighted average water potential of all nodes immediately adjacent to
840 (and distal to) the BS ("bn" stands for "bundle sheath neighbors"). Finally we defined K_{ob} as

841

842 (21) $K_{ob} = \left(K_{ox}^{-1} - K_b^{-1}\right)^{-1}$.

843

844 To allow direct comparison between measured values of K_{ox} (defined by Eqn 1) and modeled
845 values (computed by Eqn 18), we also computed alternative modeled values of K_{ox} based on the
846 volume-weighted average water potential of all tissues distal to the xylem:

847

848 (22) $K_{ox} = E_{leaf} \big/ \left|\delta\psi_{ox}\right|$,

849

850 where $\delta\psi_{ox}$ is computed in the same fashion as $\delta\psi_{ob}$, but extended to include the BS itself.
851 Modeled K_{ox} values from Eqn 18 are given in most cases in the *Results*; values from Eqn 22 are
852 used only when being compared directly to measured values (in Table II and Figure 1).

853

854 *"Unknown" parameters*

MOFLO contains six parameters that could not be estimated with the same confidence as other anatomical parameters (Table III). These are: (1) the % suppression of BS apoplastic transport by a suberized layer in BS cell walls; (2) the vertical temperature gradient within leaves, ΔT; (3) the Poiseuille radius of apoplastic nanopores, R_a; (4) the osmotic water permeability of cell membranes, P_m; (5) the ratio of true cell wall thickness to apparent thickness measured in light micrographs, ρ_{ta} (discussed further below); and (6) the ratio of the true fraction of palisade mesophyll cell area contacting horizontally adjacent cells to the apparent ratio measured in light micrographs, ρ_{fcph}. For the % suppression of BS apoplastic transport, we explored the full range of possible values (from 0 to 100%); we set the default value at 0% because there is little evidence of BS suberization in leaves of most species (Lersten, 1997). Because measured f_{cph} is most likely an overestimate (light micrographs typically cannot distinguish true horizontal connections between palisade cells and the illusion of connections created by overlap of cells in the depth plane), we set the default value for ρ_{fcph} at 0 and explored a range from 0 to 1. We used a default value of 0.1^{o}C and a range from 0 to 0.2 ^{o}C for ΔT, which spans the range of values in simulations by Rockwell et al. (2014) (note that 0.1^{o}C was the average temperature *drop* from the point of maximum temperature to the lower epidermis across species; in practice, we used the same gradient for all species, so that the absolute drop varied across species in relation to leaf thickness, as discussed earlier below Eqn 17). We explored values of R_a from 0 to 10 nm, and values of P_m from 0 to 160 μm s^{-1}, with default value of 3 nm and 40 μm s^{-1}, respectively, based on Buckley (2015).

Our anatomical measurements (John et al., 2013) suggest cell walls in our species range from 0.5 to 2.9 μm in thickness, averaging 1.4 μm across tissue types and species. These values are about five times greater than published measurements made on other species based on transmission electron microscopy (TEM) (e.g., Evans et al., 1994; Moghaddam and Wilman, 1998; Hanba et al., 2002; Scafaro et al., 2011). Light-microscopy (LM) measurements of cell wall thickness might be affected by optical artifacts (blurring near the limit of optical resolution might increase apparent wall thickness) or sampling artifacts (e.g., if a cell's perimeter is oblique to the sectioning plane, say at an angle of 45°, then the perimeter will appear at least 0.7 μm thick in a 1 μm section [~1/tan(45°)] regardless of true cell wall thickness). On the other hand, fixation for TEM requires strong dehydration that may cause cell wall shrinkage. Accurate measurement of

886 cell wall thickness is a future research direction; for the present study, we assumed by default
887 that LM measurements were overestimates by a factor of five, so the default value of ρ_{ta} was 0.2
888 and we explored a range from 0.2 to 1.0.
889
890 *Simulations to determine effects of parameters on outside-xylem hydraulic conductance*
891 MOFLO contains three classes of parameters: eight "known" biophysical parameters such as
892 molecular diffusivity and dynamic viscosity (Table I), six "unknown" parameters, discussed
893 above, that were either ambiguous in light micrographs or could not be estimated visually (Table
894 III) and 34 "known" parameters that were confidently estimated from light micrographs of
895 transverse leaf sections (Table IV). We performed three sets of simulations to explore the effects
896 of these 48 parameters on outside-xylem hydraulic conductance:
897
898 Simulation set (i). To bound the range of possible K_{ox} values consistent with the model,
899 we varied the six unknown parameters simultaneously in two simulations: one with values
900 chosen to minimise K_{ox} and another with values chosen to maximise K_{ox}. The "low-K_{ox}" values
901 were: R_a, P_m and ρ_{ta} = 50% of their respective default values, $\Delta T = 0$, $\rho_{fcph} = 0$ and 100% of BS
902 apoplastic transport blocked by a "Casparian strip". The "high-K_{ox}" values were: R_a, P_m and ρ_{ta} =
903 150% of their respective default values, $\Delta T = 0.20$ ^{o}C, $\rho_{fcph} = 1$ and no BS Casparian strip.
904
905 Simulation set (ii). To determine the mechanistic effect of each known parameter on K_{ox},
906 K_b and K_{ob}, and to distinguish between these mechanistic effects and the across-species
907 correlations between each parameter and K_{ox}, we varied each of the 34 "known" parameters
908 across the range of measured values across species, while holding the other 33 known parameters
909 at their all-species means, and holding the six "unknown" parameters at default values. These
910 parameter ranges, means and default values are given in Tables III and IV.
911
912 Simulation set (iii). To explore the importance of uncertainty in the "unknown"
913 parameters for conclusions drawn from the other simulations, we varied each of the six
914 "unknown" parameters across a range of five likely values while holding the other five unknown
915 parameters constant at their default values, and holding the 34 known parameters at their
916 measured values for each species; these simulations were repeated for each species. Embedded in

these 420 simulations (5 values × 6 unknown parameters × 14 species) are 14 (one per species) in which each unknown parameter was at its default value; these 14 simulations give the "default" predictions for each species.

Supplemental Material

Supplemental File S1. Derivation of expressions for flow area, pathlength corrections, grid areas, grid pathlengths and volume averaging basis adjustment.

Acknowledgements

TNB thanks Helen Bramley, Antonio Diaz-Espejo, John Evans and Margaret Barbour for helpful discussions.

Literature cited

Aasamaa K, Niinemets Ü, Sõber A (2005) Leaf hydraulic conductance in relation to anatomical and functional traits during Populus tremula leaf ontogeny. Tree Physiology **25:** 1409-1418

Barbour MM, Farquhar GD (2004) Do pathways of water movement and leaf anatomical dimensions allow development of gradients in H218O between veins and the sites of evaporation within leaves? Plant, Cell & Environment **27:** 107-121

Boyer JS (1985) Water transport. Annual Review of Plant Physiology **36:** 473-516

Brodribb TJ, Feild TS, Jordan GJ (2007) Leaf maximum photosynthetic rate and venation are linked by hydraulics. Plant Physiology **144:** 1890-1898

Brodribb TJ, Holbrook NM (2003) Stomatal closure during leaf dehydration, correlation with other leaf physiological traits. Plant Physiology **132:** 2166-2173

Brodribb TJ, Holbrook NM, Zwieniecki MA, Palma B (2005) Leaf hydraulic capacity in ferns, conifers and angiosperms: impacts on photosynthetic maxima. New Phytologist **165:** 839-846

Brodribb TJ, Jordan GJ (2008) Internal coordination between hydraulics and stomatal control in leaves. Plant, Cell & Environment **31:** 1557-1564

Brodribb TJ, Jordan GJ, Carpenter RJ (2013) Unified changes in cell size permit coordinated leaf evolution. New Phytologist **199:** 559-570

Buckley TN (2005) The control of stomata by water balance (Tansley Review). New Phytologist **168:** 275-292

Buckley TN (2015) The contributions of apoplastic, symplastic and gas phase pathways for water transport outside the bundle sheath in leaves. Plant, cell & environment **38:** 7-22

Buckley TN, Sack L, Gilbert ME (2011) The role of bundle sheath extensions and life form in stomatal responses to leaf water status. Plant Physiology **156:** 962-973

Carins Murphy MR, Jordan GJ, Brodribb TJ (2012) Differential leaf expansion can enable hydraulic acclimation to sun and shade. Plant, Cell & Environment **35:** 1407-1418

Carins Murphy MR, Jordan GJ, Brodribb TJ (2014) Acclimation to humidity modifies the link between leaf size and the density of veins and stomata. Plant, Cell & Environment **37:** 124-131

Chatelet DS, Wistrom CM, Purcell AH, Rost TL, Matthews MA (2011) Xylem structure of four grape varieties and 12 alternative hosts to the xylem-limited bacterium Xylella fastidious. Annals of Botany **108:** 73-85

Cochard H, Nardini A, Coll L (2004) Hydraulic architecture of leaf blades: where is the main resistance? Plant, Cell & Environment **27:** 1257-1267

Cochard H, Venisse JS, Barigah TS, Brunel N, Herbette S, Guilliot A, Tyree MT, Sakr S (2007) Putative role of aquaporins in variable hydraulic conductance of leaves in response to light. Plant Physiology **143:** 122-133

Doelger J, Rademaker H, Liesche J, Schulz A, Bohr T (2014) Diffusion and bulk flow in phloem loading: A theoretical analysis of the polymer trap mechanism for sugar transport in plants. Physical Review E **90**

Eijkel JC, Van Den Berg A (2005) Nanofluidics: what is it and what can we expect from it? Microfluidics and Nanofluidics **1:** 249-267

Evans J, Caemmerer S, Setchell B, Hudson G (1994) The Relationship Between CO₂ Transfer Conductance and Leaf Anatomy in Transgenic Tobacco With a Reduced Content of Rubisco. Functional Plant Biology **21:** 475-495

Fahlén J, Salmén L (2004) Pore and Matrix Distribution in the Fiber Wall Revealed by Atomic Force Microscopy and Image Analysis. Biomacromolecules **6:** 433-438

Farquhar GD, Raschke K (1978) On the Resistance to Transpiration of the Sites of Evaporation within the Leaf. Plant Physiology **61:** 1000-1005

Feild TS, Brodribb TJ (2013) Hydraulic tuning of vein cell microstructure in the evolution of angiosperm venation networks. New Phytologist **199:** 720-726

Fleischer A, O'Neill MA, Ehwald R (1999) The Pore Size of Non-Graminaceous Plant Cell Walls Is Rapidly Decreased by Borate Ester Cross-Linking of the Pectic Polysaccharide Rhamnogalacturonan II. Plant Physiology **121:** 829-838

Flexas J, Scoffoni C, Gago J, Sack L (2013) Leaf mesophyll conductance and leaf hydraulic conductance: an introduction to their measurement and coordination. Journal of experimental botany **64:** 3965-3981

Guyot G, Scoffoni C, Sack L (2012) Combined impacts of irradiance and dehydration on leaf hydraulic conductance: insights into vulnerability and stomatal control. Plant, cell & environment **35:** 857-871

Hanba YT, Kogami H, Terashima I (2002) The effect of growth irradiance on leaf anatomy and photosynthesis in Acer species differing in light demand. Plant, Cell & Environment **25:** 1021-1030

John GP, Scoffoni C, Sack L (2013) Allometry of cells and tissues within leaves. American journal of botany **100:** 1936-1948

Kennedy C, Šturcová A, Jarvis M, Wess T (2007) Hydration effects on spacing of primary-wall cellulose microfibrils: a small angle X-ray scattering study. Cellulose **14:** 401-408

Lersten NR (1997) Occurrence of endodermis with a casparian strip in stem and leaf. The Botanical Review **63:** 265-272

Lersten NR (1997) Occurrence of endodermis with a casparian strip in stem and leaf. Botanical Review **63:** 265-272

Martre P, Cochard H, Durand JL (2001) Hydraulic architecture and water flow in growing grass tillers (Festuca arundinacea Schreb.). Plant, Cell & Environment **24:** 65-76

McCann MC, Wells B, Roberts K (1990) Direct visualization of cross-links in the primary plant-cell wall. J Cell Sci **96:** 323-334

Meidner H (1976) Water Vapour Loss from a Physical Model of a Substomatal Cavity. Journal of Experimental Botany **27:** 691-694

Meidner H (1983) Our Understanding of Plant Water Relations: The Bidder Lecture delivered at the Hull Meeting of the Society of Experimental Biology on 7 April 1983. Journal of experimental botany**:** 1606-1618

Mertz RA, Brutnell TP (2014) Bundle sheath suberization in grass leaves: multiple barriers to characterization. Journal of Experimental Botany **65:** 3371-3380

Moghaddam PR, Wilman D (1998) Cell wall thickness and cell dimensions in plant parts of eight forage species. Journal of Agricultureal Science **131:** 59-67

Mott KA (2007) Leaf hydraulic conductivity and stomatal responses to humidity in amphistomatous leaves. Plant, Cell and Environment **30:** 1444-1449

Nardini A, Gortan E, Salleo S (2005) Hydraulic efficiency of the leaf venation system in sun- and shade-adapted species. Functional Plant Biology **32:** 953-961

Noblin X, Mahadevan L, Coomaraswamy I, Weitz DA, Holbrook NM, Zwieniecki MA (2008) Optimal vein density in artificial and real leaves. Proceedings of the National Academy of Sciences **105:** 9140-9144

Noblin X, Mahadevan L, Coomaraswamy IA, Weitz DA, Holbrook NM, Zwieniecki MA (2008) Optimal vein density in artificial and real leaves. Proceedings of the National Academy of Sciences of the United States of America **105:** 9140-9144

Pasquet-Kok J, Creese C, Sack L (2010) Turning over a new 'leaf': multiple functional significances of leaves versus phyllodes in Hawaiian Acacia koa. Plant, Cell & Environment **33:** 2084-2100

Rockwell F, Holbrook NM, Stroock AD (2014) The competition between liquid and vapor transport in transpiring leaves. Plant Physiology**:** doi: http://dx.doi.org/10.1104/pp.1114.236323

Sack L, Cowan PD, Jaikumar N, Holbrook NM (2003) The 'hydrology' of leaves: co-ordination of structure and function in temperate woody species. Plant, Cell and Environment **26:** 1343-1356

Sack L, Frole K (2006) LEAF STRUCTURAL DIVERSITY IS RELATED TO HYDRAULIC CAPACITY IN TROPICAL RAIN FOREST TREES. Ecology **87:** 483-491

Sack L, Holbrook NM (2006) Leaf hydraulics. Annual Review of Plant Biology **57:** 361-381

Sack L, Melcher PJ, Zwieniechi MA, Holbrook NM (2002) The hydraulic conductance of the angiosperm leaf lamina: a comparison of three measurement methods. Journal of Experimental Botany **53:** 2177-2184

Sack L, Scoffoni C (2013) Leaf venation: structure, function, development, evolution, ecology and applications in the past, present and future. New Phytologist **198:** 983-1000

Sack L, Scoffoni C, John GP, Poorter H, Mason CM, Mendez-Alonzo R, Donovan LA (2013) How do leaf veins influence the worldwide leaf economic spectrum? Review and synthesis. Journal of Experimental Botany **64:** 4053-4080

Sack L, Scoffoni C, John GP, Poorter H, Mason CM, Mendez-Alonzo R, Donovan LA (2014) Leaf mass per area is independent of vein length per area: avoiding pitfalls when modeling phenotypic integration. Journal of Experimental Botany **in press**

Sack L, Scoffoni C, Johnson DM, Buckley TN, Brodribb TJ (2015) The anatomical determinants of leaf hydraulic function. *In* U Hacke, ed, Functional and Ecological Xylem Anatomy. Springer International, Cham, Switzerland, pp 255-271

Sack L, Scoffoni C, McKown AD, Frole K, Rawls M, Havran JC, Tran H, Tran T (2012) Developmentally based scaling of leaf venation architecture explains global ecological patterns. Nature Communications **3:** 837

Sack L, Streeter CM, Holbrook NM (2004) Hydraulic analysis of water flow through leaves of sugar maple and red oak. Plant Physiology **134:** 1824-1833

Sack L, Tyree MT (2005) Leaf hydraulics and its implications in plant structure and function. Vascular transport in plants**:** 93-114

Scafaro AP, Von Caemmerer S, Evans JR, Atwell BJ (2011) Temperature response of mesophyll conductance in cultivated and wild Oryza species with contrasting mesophyll cell wall thickness. Plant, Cell & Environment **34:** 1999-2008

Scoffoni C, Kunkle J, Pasquet-Kok J, Vuong C, Patel AJ, Montgomery RA, Givnish TJ, Sack L (2015) Light-induced plasticity in leaf hydraulics, venation, anatomy, and gas exchange in ecologically diverse Hawaiian lobeliads. New Phytologist

Scoffoni C, McKown AD, Rawls M, Sack L (2012) Dynamics of leaf hydraulic conductance with water status: quantification and analysis of species differences under steady state. Journal of Experimental Botany **63:** 643-658

Scoffoni C, Pou A, Aasamaa K, Sack L (2008) The rapid light response of leaf hydraulic conductance: new evidence from two experimental methods. Plant, Cell and Environment **31:** 1803-1812

Scoffoni C, Sack L (2014) Are leaves "freewheelin'"? Testing for a Wheeler-type effect in leaf xylem hydraulic decline. Plant, Cell & Environment **in press**

Scoffoni C, Sack L (2015) Are leaves 'freewheelin'? Testing for a Wheeler-type effect in leaf xylem hydraulic decline. Plant, cell & environment **38:** 534-543

Simonin KA, Burns E, Choat B, Barbour MM, Dawson TE, Franks PJ (2014) Increasing leaf hydraulic conductance with transpiration rate minimizes the water potential drawdown from stem to leaf. Journal of Experimental Botany

Slewinski TL, Anderson AA, Zhang C, Turgeon R (2012) Scarecrow plays a role in establishing Kranz anatomy in maize leaves. Plant and Cell Physiology **53:** 2030-2037

Sommerville KE, Sack L, Ball MC (2012) Hydraulic conductance of Acacia phyllodes (foliage) is driven by primary nerve (vein) conductance and density. Plant, cell & environment **35:** 158-168

Sperry JS (2000) Hydraulic constraints on gas exchange. Agricultural and Forest Meteorology **104:** 13-23

Tyree MT, Cochard H (2003) Vessel contents of leaves after excision: a test of the Scholander assumption. Journal of Experimental Botany **54:** 2133-2139

Tyree MT, Yianoulis P (1980) The site of water evaporation from sub-stomatal cavities, liquid path resistances and hydroactive stomatal closure. Annals of Botany **45:** 175-193

Voicu MC, Cooke JE, Zwiazek JJ (2009) Aquaporin gene expression and apoplastic water flow in bur oak (Quercus macrocarpa) leaves in relation to the light response of leaf hydraulic conductance. Journal of Experimental Botany**:** erp239

Wylie RB (1952) The Bundle Sheath Extension in Leaves of Dicotyledons. American Journal of Botany **39:** 645-651

Yang S, Tyree MT (1994) Hydraulic architecture of Acer saccharum and A. rubrum: comparison of branches to whole trees and the contribution of leaves to hydraulic resistance. Journal of Experimental Botany **45:** 179-186

Zsögön A, Alves Negrini AC, Peres LEP, Nguyen HT, Ball MC (2015) A mutation that eliminates bundle sheath extensions reduces leaf hydraulic conductance, stomatal conductance and assimilation rates in tomato (Solanum lycopersicum). New Phytologist **205:** 618-626

Zwieniecki MA, Boyce CK (2014) Evolution of a unique anatomical precision in angiosperm leaf venation lifts constraints on vascular plant ecology. Proceedings of the Royal Society B-Biological Sciences **281**

Figure Legends

Figure 1. Comparison of observed K_{ox} (red bars, means ± SEs) with ranges of simulated K_{ox} values possible for wide variation in "unknown" parameters ("simulated (low)" to "simulated (high)"; grey bars). For three of the unknown parameters (Poiseuille radius of apoplastic bulk flow pathways (R_a), membrane permeability (P_m) and the ratio of cell wall thickness used in the simulation to values measured by light microscopy (ρ_{ta})), the "high" and "low" K_{ox} simulations used default values plus or minus 50%, respectively. For the other three unknown parameters (BS Casparian strip, horizontal palisade contact fractions (ρ_{fcph}) and vertical temperature gradient (ΔT)), we used the upper or lower limits of possible values (0 or 100% of apoplastic transport was blocked by a Casparian strip; $\rho_{fcph} = 0$ or 1; and $\Delta T = 0$ or 0.2^{o}C). Default values for these parameters are given in Table III.

Figure 2. Example of the distribution of water potential drawdown relative to the xylem ($\delta\psi$) in a transverse leaf section as computed by MOFLO (for *Comarostaphylos diversifolia* in this example). Water potential is greatest in the bundle sheath (lower left edge) and most negative in the lower epidermis near the areole center (lower right). A large region of palisade mesophyll (top center) remains at quite high water potential, because most water flow and thus most potential drawdown occurs through the spongy mesophyll (lower center). The volume-weighted average water potential drawdown ($\delta\psi_{ox}$) in this example is -0.60 MPa, or -0.63 MPa excluding the bundle sheath itself ($\delta\psi_{ob}$), but the largest drawdown ($\delta\psi_{min}$) is -2.23 MPa, and the volume-weighted mean drawdown to the lower epidermis is -1.02 MPa.

Figure 3. Bulk hydraulic conductivities (water flow per unit bulk area, per unit water potential gradient) for different tissue types (vertical axis) and flow pathways (colors). All-species means are shown by the bars, and total bulk conductivities across all pathways for each of 14 species are shown by open symbols. Colors: grey = apoplastic (apo); white = transmembrane (tm); black = gas phase. Tissue types: epid = epidermis; BSE = bundle sheath extension; BS = bundle sheath; pal = palisade mesophyll; spo = spongy mesophyll; L = lower; U = upper; V = vertical transport; H = horizontal transport.

Figure 4. Effects of Poiseuille radius of apoplastic nanopores (R_a, horizontal axes) and osmotic water permeability of membranes (P_m, vertical axes) on outside-xylem hydraulic conductance (K_{ox}, contours) for two sets of conditions: A, default values for all parameters, as given in Tables III and IV, including no blockage of apoplastic transport in the bundle sheath; B, default values for all parameters except that apoplastic pathways in the bundle sheath are blocked by a suberized layer or "Casparian strip". Grey symbols: location of default values for R_a and P_m (3 nm and 40 μm s^{-1}, respectively).

Figure 5. Effect of variation in the assumed values of anatomical parameters that are difficult to estimate with confidence by light microscopy (the fraction of horizontal surface area on palisade cells that is in contact with adjacent palisade cells, "horz palisade contact fraction" or f_{cph}; and cell wall thicknesses), expressed as ratios of the values used in simulations to values measured by microscopy. Asterices indicate default values for these ratios (zero for f_{cph}, and 0.2 for cell wall thicknesses, respectively).

Figure 6. Effect of variation in vertical temperature gradients within leaves on outside-xylem hydraulic conductance, K_{ox} (thick grey line), and the components of K_{ox} that are attributable to gas phase transport that is independent of vertical gradients ("isothermal gas", dash-dot line), gas phase transport that is driven by vertical temperature gradients independent of water potential gradients ("anisothermal gas", dashed line) and the sum of those two gas phase components ("total gas", solid black line).

Figure 7. Effects of leaf temperature on outside-xylem hydraulic conductance, K_{ox}, and its components outside the bundle sheath (K_{ob}) and across the bundle sheath (K_b, right-hand axis). Note that K_b is plotted on a different scale than K_{ob} and K_{ox}.

Figure 8. Effects of Poiseuille radius of apoplastic nanopores (R_a, horizontal axes) and osmotic water permeability of membranes (P_m, vertical axes) on percent of outside-xylem hydraulic resistance contributed by the bundle sheath (% resistance in BS, contours) for two sets of conditions: A, default values for all parameters, as given in Tables III-IV, including no blockage of apoplastic transport in the bundle sheath; B, default values for all parameters except that apoplastic pathways in the bundle sheath are blocked by a suberized layer or "Casparian strip". Grey symbols: location of default values for R_a and P_m (3 nm and 40 μm s^{-1}, respectively).

Figure 9. Effects of vein length per unit leaf area (VLA) on A, hydraulic conductance across the bundle sheath, K_b; B, outside-bundle sheath hydraulic conductance, K_{ob}, and C, outside-xylem hydraulic conductance, K_{ox}. Solid lines are mechanistic relationships obtained by varying VLA in the model while holding all other parameters constant. Open symbols are values for each of 14 species, and dashed lines are the cross-species correlations between VLA and each conductance. r^2 and p values shown are for the cross-species correlations.

Figure 10. Effects of two anatomical parameters related to vertical distance between veins and the epidermis on outside-xylem hydraulic conductance, K_{ox}: A, the ratio of the distance between the bundle sheath [BS] and the upper epidermis to the distance to the lower epidermis, and B, total leaf thickness. Solid lines are mechanistic relationships obtained by varying the parameters shown on the x-axes in the model while holding all other parameters constant. Open symbols are values for each of 14 species, and dashed lines are the cross-species correlations between the parameters on the x-axes and each conductance. r^2 and p values shown are for the cross-species correlations.

Figure 11. Effects of four anatomical parameters related to spongy mesophyll anatomy on outside-xylem hydraulic conductance, K_{ox}: A, spongy mesophyll cell radius, B, spongy mesophyll tissue airspace fraction, C, the fraction of spongy mesophyll surface area that is in contact with adjacent spongy cells, and D, the ratio of palisade tissue thickness to spongy tissue thickness. Solid lines are mechanistic relationships obtained by varying the parameters shown on the x-axes in the model while holding all other parameters constant. Open symbols are values for each of 14 species, and dashed lines are the cross-species correlations between the parameters on the x-axes and each conductance. r^2 and p values shown are for the cross-species correlations.

Figure 12. Differences in outside-xylem hydraulic conductance, K_{ox}, computed by the model for homobaric (white bar) and heterobaric species (grey bars). For heterobaric species, an additional simulation was performed in which nodes in the grid corresponding to bundle sheath extensions (BSEs) were assigned conductivities corresponding to the adjacent mesophyll (grey bar with hash marks; "heterobaric –BSEs").

Figure 13. Simulation of the effect of increasing transpiration rate (E) three-fold at the upper surface of an amphistomatous leaf (*Helianthus annuus*) while holding it constant at the lower surface. Color contours represent water potential drawdowns from the xylem ($\delta\psi$); the areole margin (minor vein) is at left and the areole center is at right; the upper and lower surfaces are at top and bottom, respectively. Left panel: before the change in transpiration rate; right panel: after the change. Initial and final transpiration rates for both surfaces, and $\delta\psi$ values for the epidermal nodes at the center of the areole at both surfaces, are shown at bottom. These simulations used default values for all parameters as given in Tables III-IV.

Figure 14. Diagram of MOFLO structure. The model represents an areole (the smallest region of a leaf bounded by minor veins) as a circular and radially-symmetrical region (top and middle right in figure). Flow is simulated through a grid with 744 nodes (24 columns and 31 rows; bottom left) representing a transverse section through the areole; the left-hand edge of this grid corresponds to the outer margin of the areole, and the right-hand edge corresponds to the center of the areole. Grid nodes are allocated to tissue types based on measured tissue dimensions (cf. grid at bottom left and diagram of tissue types at bottom right).

Table I. List of parameters.

Variables and parameters referred to in this study, other than leaf anatomical parameters (Table IV) or unknown parameters (Table III) , with symbols and units. (1): volume-weighted average. BS: bundle sheath. $D_{wa} = 2.178 \cdot 10^{-5} \cdot (T/273.15)^{1.81}$ m^2 s^{-1}, $D_{ww} = 1.635 \cdot 10^{-8} \cdot (T/215.05 - 1)^{2.063}$ m^2 s^{-1}, and $\eta = 1.95 \cdot 10^{14} \cdot T^{-7}$ Pa s, where T is in kelvins; $R_{gas} = 8.314462$ Pa m^3 mol^{-1} K^{-1}; $v_w = 1.8 \cdot 10^{-5}$ m^3 mol^{-1}. $T = 25^{o}$C or 298.15 K except where otherwise noted.

variable or parameter	symbol	units
bulk area for flow between two nodes	a	m^2
bulk area for flow between nodes j and i	a_{ji}	m^2
ratio of actual flow pathlength to simple (direct) pathlength	β	unitless
molecular diffusivity of water vapour in air	D_{wa}	m^2 s^{-1}
molecular diffusivity of water in liquid water	D_{wa}	m^2 s^{-1}
vertical temperature difference from T_{max} to lower surface	ΔT	K
vector of water potential decreases relative to the xylem	$\boldsymbol{\delta\psi}$	MPa
water potential drawdown to BS neighbors (1)	$\delta\psi_{bn}$	MPa
water potential drawdown to node i	$\delta\psi_i$	MPa
water potential drawdown to all nodes outside the BS (1)	$\delta\psi_{ob}$	MPa
vector of E_i minus $\Sigma_j\{F_{aniso,ji}\}$	$\mathbf{e}$	mol s^{-1}
transpiration rate from node i	E_i	mol s^{-1}
element of **e** for node i	e_i	mol s^{-1}
leaf level transpiration rate	E_{leaf}	mmol m^{-2} s^{-1}
anisothermal gas phase flux into node i	$F_{aniso,ji}$	mol s^{-1}
xylem fraction of total leaf water	f_x	unitless
ratio of actual flow area to simple (bulk) flow area	γ	unitless
dynamic viscosity of water	η	Pa s
matrix of internodal conductances	$\mathbf{K}$	mol s^{-1} MPa^{-1}
conductivity	κ	mol s^{-1} (MPa m^{-1})$^{-1}$ m^{-2}
conductance for anisothermal gas phase flow from node j to i	$K_{aniso,ji}$	mol s^{-1} MPa^{-1}
BS conductance	K_b	mmol m^{-2} s^{-1} MPa^{-1}
conductance for isothermal gas phase flow from node j to i	$K_{iso,ji}$	mol s^{-1} MPa^{-1}
conductance between node j and a BS node (b)	K_{jb}	mol s^{-1} MPa^{-1}
conductance between nodes j and i	K_{ji}	mol s^{-1} MPa^{-1}
leaf hydraulic conductance	K_{leaf}	mmol m^{-2} s^{-1} MPa^{-1}
outside-BS hydraulic conductance	K_{ob}	mmol m^{-2} s^{-1} MPa^{-1}
outside-xylem hydraulic conductance	K_{ox}	mmol m^{-2} s^{-1} MPa^{-1}
xylem hydraulic conductance	K_x	mmol m^{-2} s^{-1} MPa^{-1}
conductance between xylem and a BS node (b)	K_{xb}	mol s^{-1} MPa^{-1}
transcellular pathlength	L_c	m
direct pathlength between two nodes	l	m
direct pathlength between nodes j and i	l_{ji}	m
saturation vapour pressure at nodes i or j	$p_{sat,i}$, $p_{sat,j}$	Pa
ideal gas constant	R_{gas}	Pa m^3 mol^{-1} K^{-1}
outside-xylem hydraulic resistance ($1/K_{ox}$)	R_{ox}	(mmol m^{-2} s^{-1} MPa^{-1})$^{-1}$
temperature as a function of relative depth in leaf (x)	$T(z)$	K
temperature at nodes i or j	T_i, T_j	K
maximum temperature in leaf	T_{max}	K
volume of water in node i	v_i	m^3
molar volume of liquid water	v_w	m^3 mol^{-1}
water potential; water potentials at nodes i or j	ψ, ψ_i, ψ_j	MPa
relative depth in leaf (0 = top)	z	unitless
relative depth in leaf at which temperature is greatest	z_{max}	unitless

Table II. Measured vs. modeled outside-xylem hydraulic conductance.
Measured and modeled outside-xylem hydraulic conductance (K_{ox}) for eight species. *Modeled values are based on default simulation conditions and calculated using Eqn 18 (based on average outside-xylem water potential rather than outside-BS water potential). Units for all conductances are mmol m^{-2} s^{-1} MPa^{-1}.

	K_{ox}	
species	**measured**	**modeled***
Cercocarpus betuloides	4.7	30.2
Comarostophylos diversifolia	3.5	8.3
Helianthus annuus	7.5	4.9
Hedera canariensis	6.1	11.8
Lantana camara	11.3	26.8
Magnolia grandiflora	3.7	15.4
Quercus agrifolia	4.5	30.7
Salvia canariensis	54.3	5.8
mean ± SE	11.9 ± 6.1	16.8 ± 3.9
median ± MAD	5.4 ± 1.8	13.6 ± 8.2

Table III. "Unknown" parameters.

Parameters that could not be estimated with confidence from light micrographs across species (these are referred to in the text as "unknown" parameters), with ranges and default values used in simulations. "BS" means "bundle sheath"; "pal horz" means "palisade horizontal".

parameter	symbol	range	default	units
% of BS apoplastic transport blocked by suberized layer	-	0-100	0	%
membrane permeability	P_m	0-160	40	$\mu m\ s^{-1}$
effective Poiseuille radius of cell wall	R_a	0-10	3	nm
ratio of true pal horz connectivity to value from microscopy	ρ_{fcph}	0-1.0	0	-
ratio of true cell wall thicknesses to values from microscopy	ρ_{ta}	0.2-1.0	0.2	-
vertical temperature gradient in leaf	ΔT	0-0.2	0.1	^{o}C

Table IV. Anatomical parameter values.

Anatomical parameter values measured for 14 species. Species codes are: BAGA, *Bauhinia galpinii*; CASA, *Camellia sasanqua*; CEBE, *Cercocarpus betuloides*; CODI, *Comarostophylos diversifolia*; HEAN, *Helianthus annuus*; HEAR, *Heteromeles arbutifolia*; HECA, *Hedera canariensis*; LACA, *Lantana camara*; MAGR, *Magnolia grandiflora*; PLRA, *Platanus racemosa*; QUAG, *Quercus agrifolia*; RAIN, *Raphiolepis indica*; ROCO, *Romneya coulterii*, SACA, *Salvia canariensis*.

			species													
parameter	symbol	units	BAGA	CASA	CEBE	CODI	HEAN	HEAR	HECA	LACA	MAGR	PLRA	QUAG	RAIN	ROCO	SACA
cell wall thicknesses																
bundle sheath cell wall thickness	t_{abs}	µm	0.56	0.99	0.69	0.73	0.63	1.71	1.21	0.83	1.14	0.79	1.02	1.14	0.82	0.75
epidermal cell wall thickness (lower)	t_{ael}	µm	0.63	2.54	1.74	2.03	0.80	1.80	1.84	1.55	2.40	1.48	1.80	2.10	1.96	1.22
epidermal cell wall thickness (upper)	t_{aeu}	µm	0.95	2.93	2.24	2.67	0.80	1.76	1.98	1.56	2.30	1.66	1.97	1.94	2.04	1.42
palisade cell wall thickness	t_{ap}	µm	0.54	1.48	1.08	1.41	0.66	1.18	1.48	1.06	1.73	0.81	1.23	1.17	1.36	0.93
spongy cell wall thickness	t_{as}	µm	0.64	2.15	1.30	1.23	0.54	1.37	1.77	1.09	1.76	0.81	1.53	1.92		0.91
BSE cell wall thickness	t_{ax}	µm	1.07		1.42	1.26	1.54				2.30	0.69	1.34		1.34	
cell scale parameters																
palisade cell height	h_p	µm	27.9	69.4	29.3	47.4	48.1	43.6	45.3	39.8	60.8	50.9	35.0	47.0	36.6	34.2
bundle sheath cell perimeter	p_{bsc}	µm	28.3	69.6	40.1	46.7	55.5	69.7	80.5	59.8	66.4	47.6	47.0	73.2	58.3	37.6
palisade radius	r_p	µm	6.7	20.9	8.0	14.1	14.4	10.5	26.8	11.7	21.6	11.7	8.7	11.5	12.6	12.4
spongy radius	r_s	µm	9.0	27.8	6.0	19.7	17.2	22.0	25.0	14.6	24.6	11.1	10.4	25.5		11.3
width of upper epidermal cell	w_{el}	µm	11.2	25.0	9.4	11.1	19.2	17.6	21.7	14.0	19.9	18.4	11.1	14.1	41.7	13.6
width of lower epidermal cell	w_{eu}	µm	16.4	12.5	18.1	15.6	14.9	21.5	10.5	16.4	18.0	18.4	18.7	39.6	42.0	16.2
width of one BSE cell	w_x	µm	8.2		19.9	18.6	34.3				23.1	9.4	16.3		28.5	
tissue scale parameters																
distance from BS to lower epid	h_{xltot}	µm	7.7	111.5	47.9	73.9	52.7	97.7	121.5	30.3	145.0	39.2	41.0	195.1	76.0	41.7
distance from BS to upper epid	h_{xutot}	µm	29.8	94.3	113.8	140.9	70.9	92.9	112.1	77.3	220.2	76.2	140.3	126.8	92.3	65.4
total perimeter of vascular bundle	p_{bs}	µm	143.9	391.3	247.1	300.5	195.7	525.2	288.0	273.0	343.6	194.7	282.3	399.1	359.5	185.2
lower epidermis thickness	t_{el}	µm	9.5	13.1	18.9	8.5	11.3	17.6	9.2	10.4	10.1	11.1	12.5	13.9	34.1	8.9
upper epidermis thickness	t_{eu}	µm	16.0	13.9	19.0	14.8	13.3	18.9	11.0	18.4	47.4	17.9	19.1	36.8	40.4	16.2
palisade thickness	t_p	µm	27.6	121.9	97.6	100.8	67.2	95.1	66.1	85.7	195.4	72.4	118.9	107.2	294.3	87.0
spongy thickness	t_s	µm	37.5	259.0	112.5	160.6	90.6	236.4	215.5	93.3	268.2	93.5	127.5	304.5		66.2
total width of BSE	w_{xtot}	µm	15.3		21.1	31.2	13.2				40.4	6.2	26.4		17.7	
dimensionless parameters																
palisade horizontal connectivity	f_{cph}	-	0.42	0.22	0.18	0.22	0.22	0.07	0.03	0.21	0.52	0.49	0.74	0.85	0.57	0.60
palisade vertical connectivity	f_{cpv}	-	0.44	0.49	0.35	0.58	0.42	0.64	0.64	0.62	0.43	0.28	0.36	0.24	0.24	0.33
spongy mesophyll connectivity	f_{cs}	-	0.31	0.50	0.17	0.21	0.23	0.32	0.28	0.17	0.23	0.28	0.23	0.24		0.23
leaf airspace fraction in palisade	p_p	-	0.10	0.20	0.12	0.10	0.27	0.23	0.13	0.18	0.18	0.40	0.07	0.12	0.35	0.20
leaf airspace fraction in spongy	p_s	-	0.10	0.42	0.63	0.40	0.43	0.60	0.52	0.33	0.32	0.45	0.27	0.40		0.27
leaf scale parameters																
vein length per unit area	VLA	mm^{-1}	4.98	3.31	7.74	4.17	9.32	4.63	3.00	9.75	5.16	4.97	7.30	3.90	4.15	4.15

Table V. Simulated outside-xylem hydraulic conductance and its components.
Values of outside-xylem hydraulic conductance (K_{ox}) and its components, bundle sheath and outside-bundle sheath conductances (K_b and K_{ob}, respectively), simulated under default values for all parameters as given in Tables III-IV (left three columns of results). Simulated K_{ox} values for three additional scenarios involving "unknown" parameters are shown in the right three columns of results: the presence/absence of a Casparian strip in the bundle sheath ("Casp"), specification of cell wall thicknesses in the model at 20% or 100% of values measured by light microscopy (ρ_{ta} = 0.2 or 1.0), and specification of horizontal palisade cell connectivity (contact fraction) at 0% or 100% of values measured by light microscopy (ρ_{fcph} = 0 or 1). Means ± SEs and medians ± median absolute deviations are shown at bottom. Units: mmol m^{-2} s^{-1} MPa^{-1}.

Casp	*no*			**yes**	no	no
ρ_{ta}	*0.2*			0.2	**1.0**	0.2
ρ_{fcph}	*0*			0	0	**1**
species	K_b	K_{ob}	K_{ox}		K_{ox}	
Bauhinia galpinii	24.9	4.8	4.0	2.1	16.8	4.7
Camellia sasanqua	33.6	13.0	9.4	5.2	29.5	9.7
Cercocarpus betuloides	98.5	38.1	27.5	10.5	109.4	27.7
Comarostophylos diversifolia	19.1	13.4	7.9	4.3	27.1	8.0
Helianthus annuus	35.8	14.4	10.3	5.7	29.7	10.5
Heteromeles arbutifolia	90.8	11.9	10.5	7.1	26.8	10.5
Hedera canariensis	30.4	5.2	4.5	2.8	13.2	4.5
Lantana camara	136.3	25.9	21.7	10.4	80.0	22.2
Magnolia grandiflora	112.9	16.3	14.3	6.7	53.2	14.4
Platanus racemosa	45.4	10.7	8.7	4.4	29.9	9.4
Quercus agrifolia	72.0	47.4	28.6	9.3	124.7	29.6
Raphiolepis indica	7.1	17.0	5.0	3.6	15.6	5.3
Romneya coulterii	66.7	9.2	8.1	5.1	31.9	11.2
Salvia canariensis	37.7	6.4	5.5	2.7	21.9	7.7
mean ± SE	58±10	17±3	11.8±2.2	5.7±0.7	44±10	12.5±2.2
median ± MAD	42±24	13±4	9±3.8	5.1±1.8	30±10	10.1±3.4

Table VI. Mechanistic trait analysis.

Standardised slopes, expressed as percents, for relationships between leaf anatomical traits and outside-xylem hydraulic conductance (K_{ox}). "mech" values are mechanistic effects, i.e., relationships obtained by varying individual parameters in isolation, whereas "corr" values are for correlations between parameters and K_{ox} across species. "sig (corr)" is the significance of across-species correlations (***: $p < 0.001$; **: $p < 0.01$; *: $p < 0.05$; ns: $p > 0.05$). r^2 (corr) is the correlation coefficient for "corr" slopes. For palisade and spongy thicknesses, mechanistic effects were assessed using palisade/spongy thickness ratio; likewise, mechanistic effects for distances between the BS and epidermis were assessed using the ratio of distances above/below the BS.

parameter	symbol	slope (mech)	slope (corr)	sig (corr)	r^2 (corr)
cell wall thicknesses					
bundle sheath cell wall thickness	t_{abs}	15	-8	ns	0.00
epidermal cell wall thickness (lower)	t_{ael}	0	30	ns	0.01
epidermal cell wall thickness (upper)	t_{aeu}	0	37	ns	0.02
palisade cell wall thickness	t_{ap}	6	23	ns	0.00
spongy cell wall thickness	t_{as}	36	13	ns	0.00
BSE cell wall thickness	t_{ax}	13	51	*	0.06
cell scale parameters					
height of individual palisade cell	h_p	-5	-64	*	0.05
bundle sheath cell perimeter	p_{bsc}	-32	-47	ns	0.03
palisade radius	r_p	-2	-55	**	0.10
spongy radius	r_s	-30	-73	***	0.19
width of epidermal cells (lower)	w_{el}	0	-48	**	0.10
width of epidermal cells (upper)	w_{eu}	0	-21	ns	0.01
width or height of one BSE cell	w_x	-12	6	ns	0.00
tissue scale parameters					
distance from BS to lower epidermis	h_{xltot}		-31	**	0.09
distance from BS to upper epidermis	h_{xutot}		48	**	0.09
total perimeter of vascular bundle	p_{bs}	22	-3	ns	0.00
epid thickness lower	t_{el}	-18	13	ns	0.00
epid thickness upper	t_{eu}	11	1	ns	0.00
palisade thickness	t_p		10	ns	0.00
spongy thickness	t_s		-22	ns	0.03
total width of BSE	w_{xtot}	14	39	**	0.08
dimensionless parameters					
max palisade horz connectivity	f_{cph}	0	-1	ns	0.00
palisade vertical connectivity	f_{cpv}	0	-1	ns	0.00
spongy mesophyll connectivity	f_{cs}	24	-90	***	0.17
leaf airspace fraction in palisade	p_p	3	-37	**	0.07
leaf airspace fraction in spongy	p_s	22	31	ns	0.02
ratio of palisade to spongy thickness	-	-4	67	***	0.14
ratio of distances above/below BS	-	34	59	***	0.21
leaf scale parameters					
total leaf thickness	-	-18	-4	ns	0.00
vein length per unit area	VLA	110	121	***	0.47

Table VII. Summary data for species used in this study.

Species codes are: BAGA, *Bauhinia galpinii*; CASA, *Camellia sasanqua*; CEBE, *Cercocarpus betuloides*; CODI, *Comarostophylos diversifolia*; HEAN, *Helianthus annuus*; HEAR, *Heteromeles arbutifolia*; HECA, *Hedera canariensis*; LACA, *Lantana camara*; MAGR, *Magnolia grandiflora*; PLRA, *Platanus racemosa*; QUAG, *Quercus agrifolia*; RAIN, *Raphiolepis indica*; ROCO, *Romneya coulterii*, SACA, *Salvia canariensis*. LH, leaf habit (e:evergreen, d:deciduous); LF: life form (t:tree, s:shrub, ah:annual herb, l:liana, ps:perennial shrub, ph:perennial herb), HE/HO (HE:heterobaric, HO:homobaric), LA, leaf area (cm^2), LMA, leaf mass per unit area ($g\ m^{-2}$), TLP, osmotic pressure at turgor loss (MPa).

species	family	origin	LH	LF	HE/HO	LA	LMA	TLP
BAGA	Fabaceae	Africa	e	t	HE	33.1 ± 4.3	45.0 ± 1.6	1.41 ± 0.07
CASA	Theaceae	Japan	e	s	HO	14.0 ± 1.5	178 ± 9	2.12 ± 0.18
CEBE	Rosaceae	Calif, Mexico	e	s	HE	7.0 ± 2.1	31.2 ± 1.1	1.09 ± 0.12
CODI	Ericaceae	Calif, Mexico	e	s	HE	11.1 ± 0.4	61.4 ± 4.2	1.37 ± 0.04
HEAN	Asteraceae	N America	d	ah	HE	95.0 ± 8.9	220 ± 11	2.06 ± 0.05
HEAR	Araliaceae	Calif, Mexico	e	l	HO	107 ± 15	56.3 ± 2.3	1.19 ± 0.09
HECA	Rosaceae	Canary Islands	e	s	HO	30.5 ± 3.7	211 ± 8.3	2.07 ± 0.11
LACA	Verbenaceae	Pantropical	d	ps	HO	31.6 ± 1.6	121 ± 23	2.59 ± 0.03
MAGR	Magnoliaceae	Southern US	e	t	HE	118 ± 20	253 ± 17	3.45 ± 0.34
PLRA	Platanaceae	Calif, Mexico	d	t	HE	67 ± 38	84.1 ±11.0	1.98 ± 0.09
QUAG	Fagaceae	Calif, Mexico	e	t	HE	35 ± 10	185 ± 12	2.53 ± 0.10
RAIN	Rosaceae	S China, India	e	s	HO	137 ± 90	188 ± 8	3.00 ± 0.12
ROCO	Papaveraceae	Calif, Mexico	d	ph	HE	24.0 ± 9.7	78.1 ± 3.7	1.40 ± 0.07
SACA	Lamiaceae	Canary Islands	d	ph	HO	19.1 ± 3.4	41.4 ± 6.0	1.18 ± 0.07

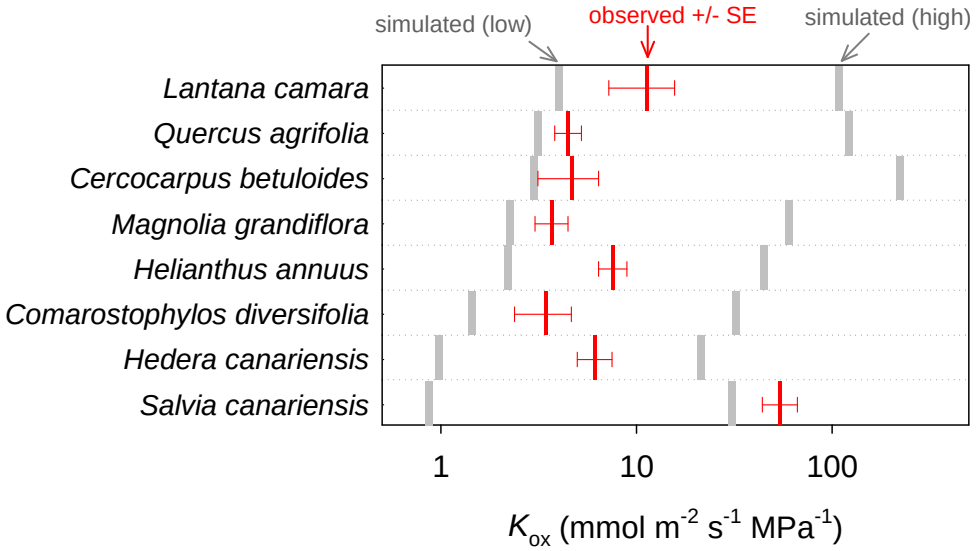


Figure 1. Comparison of observed K_{ox} (red bars, means ± SEs) with ranges of simulated K_{ox} values possible for wide variation in "unknown" parameters ("simulated (low)" to "simulated (high)"; grey bars). For three of the unknown parameters (Poiseuille radius of apoplastic bulk flow pathways (R_a), membrane permeability (P_m) and the ratio of cell wall thickness used in the simulation to values measured by light microscopy (ρ_{ta})), the "high" and "low" K_{ox} simulations used default values plus or minus 50%, respectively. For the other three unknown parameters (BS Casparian strip, horizontal palisade contact fractions (ρ_{fcph}) and vertical temperature gradient (ΔT)), we used the upper or lower limits of possible values (0 or 100% of apoplastic transport was blocked by a Casparian strip; ρ_{fcph} = 0 or 1; and ΔT = 0 or 0.2°C). Default values for these parameters are given in Table III.

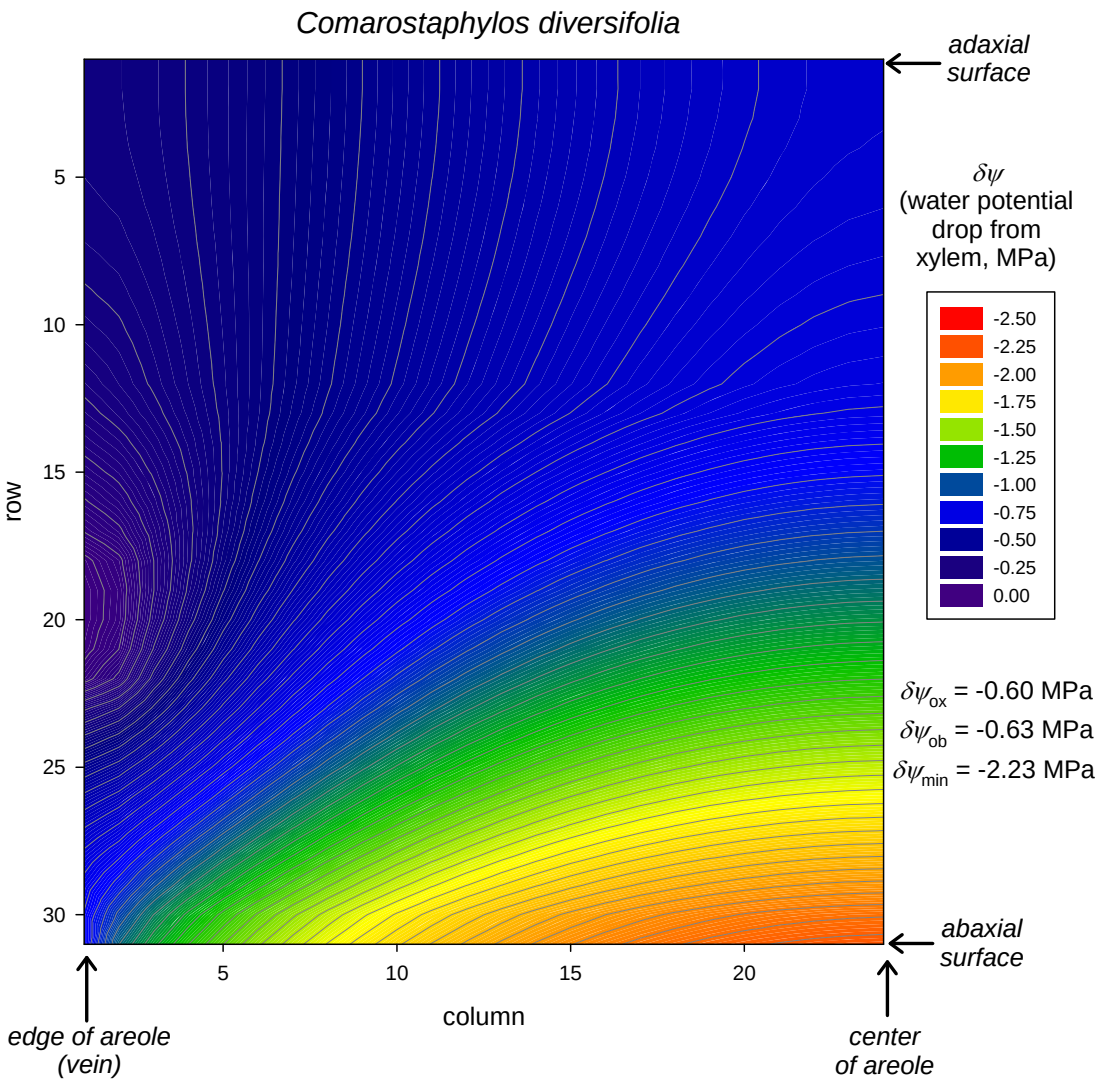


Figure 2. Example of the distribution of water potential drawdown relative to the xylem ($\delta\psi$) in a transverse leaf section as computed by MOFLO (for *Comarostaphylos diversifolia* in this example). Water potential is greatest in the bundle sheath (lower left edge) and most negative in the lower epidermis near the areole center (lower right). A large region of palisade mesophyll (top center) remains at quite high water potential, because most water flow and thus most potential drawdown occurs through the spongy mesophyll (lower center). The volume-weighted average water potential drawdown ($\delta\psi_{ox}$) in this example is -0.60 MPa, or -0.63 MPa excluding the bundle sheath itself ($\delta\psi_{ob}$), but the largest drawdown ($\delta\psi_{min}$) is -2.23 MPa, and the volume-weighted mean drawdown to the lower epidermis is -1.02 MPa.

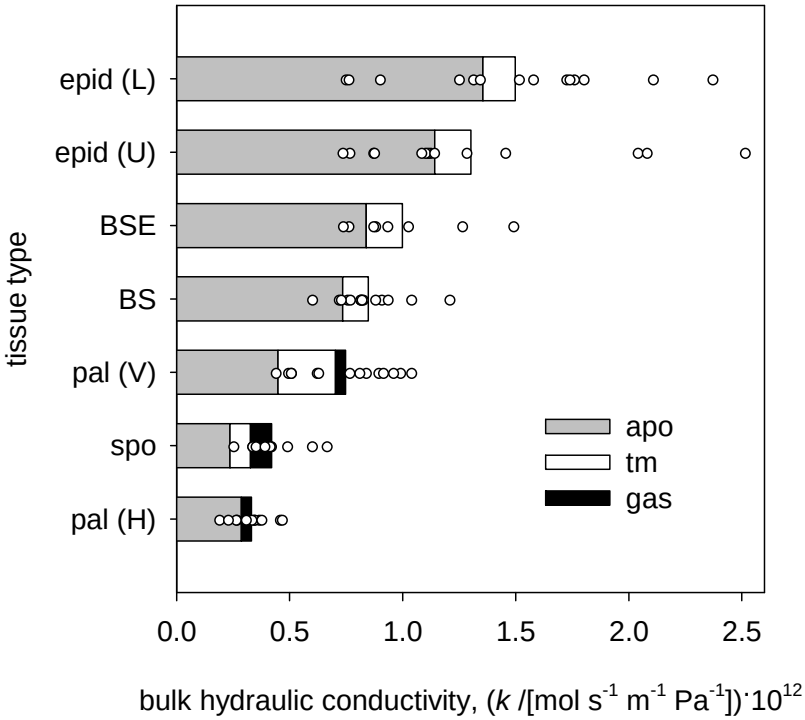


Figure 3. Bulk hydraulic conductivities (water flow per unit bulk area, per unit water potential gradient) for different tissue types (vertical axis) and flow pathways (colors). All-species means are shown by the bars, and total bulk conductivities across all pathways for each of 14 species are shown by open symbols. Colors: grey = apoplastic (apo); white = transmembrane (tm); black = gas phase. Tissue types: epid = epidermis; BSE = bundle sheath extension; BS = bundle sheath; pal = palisade mesophyll; spo = spongy mesophyll; L = lower; U = upper; V = vertical transport; H = horizontal transport.

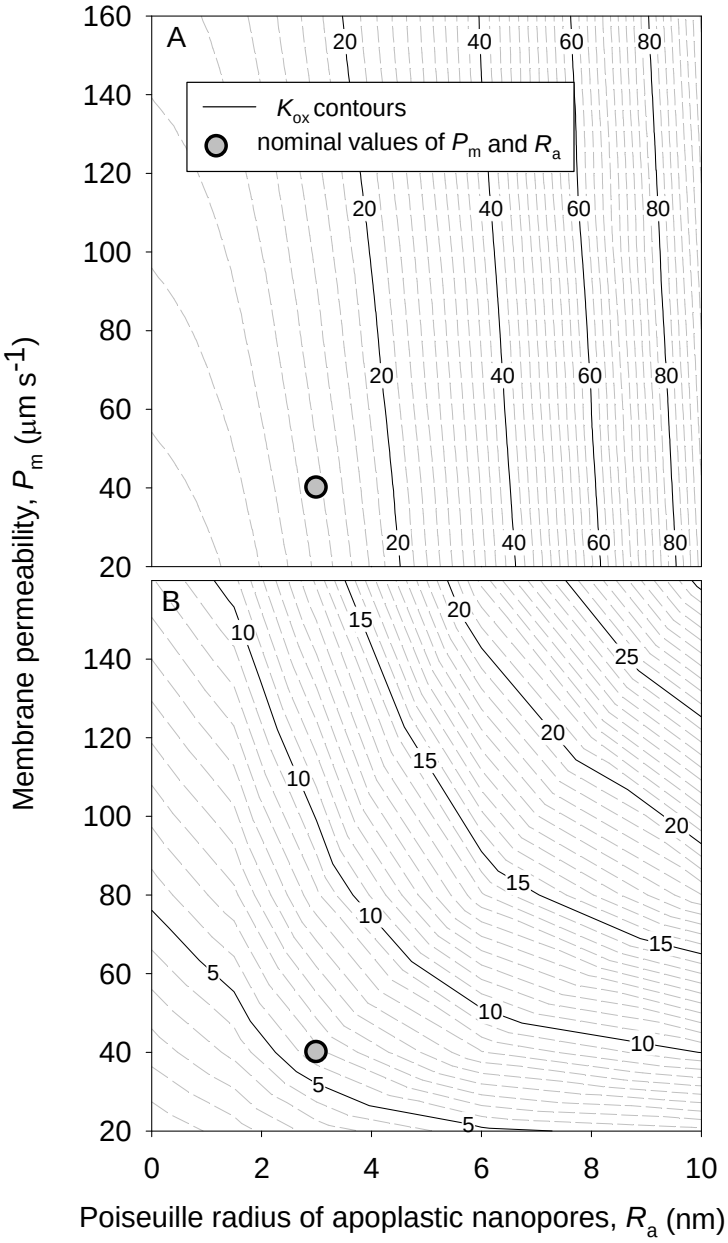


Figure 4. Effects of Poiseuille radius of apoplastic nanopores (R_a, horizontal axes) and osmotic water permeability of membranes (P_m, vertical axes) on outside-xylem hydraulic conductance (K_{ox}, contours) for two sets of conditions: (a) default values for all parameters, as given in Tables III and IV, including no blockage of apoplastic transport in the bundle sheath; (b) default values for all parameters except that apoplastic pathways in the bundle sheath are blocked by a suberized layer or "Casparian strip". Grey symbols: location of default values for R_a and P_m (3 nm and 40 μm s^{-1}, respectively).

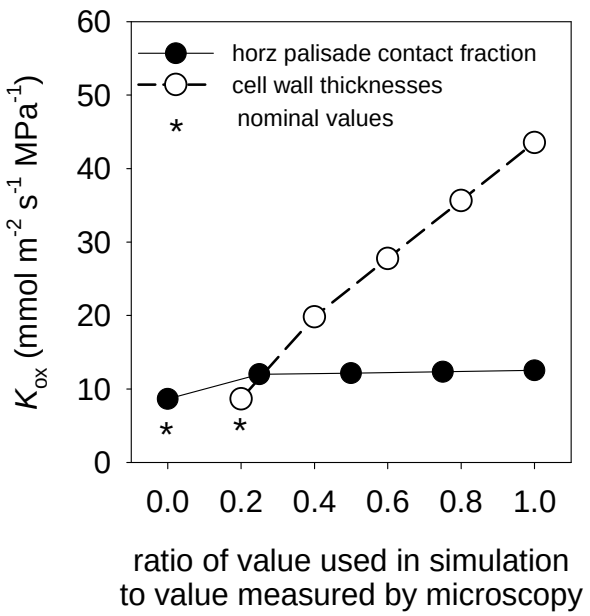


Figure 5. Effect of variation in the assumed values of anatomical parameters that are difficult to estimate with confidence by light microscopy (the fraction of horizontal surface area on palisade cells that is in contact with adjacent palisade cells, "horz palisade contact fraction" or f_{cph}; and cell wall thicknesses), expressed as ratios of the values used in simulations to values measured by microscopy. Asterices indicate default values for these ratios (zero for f_{cph}, and 0.2 for cell wall thicknesses, respectively).

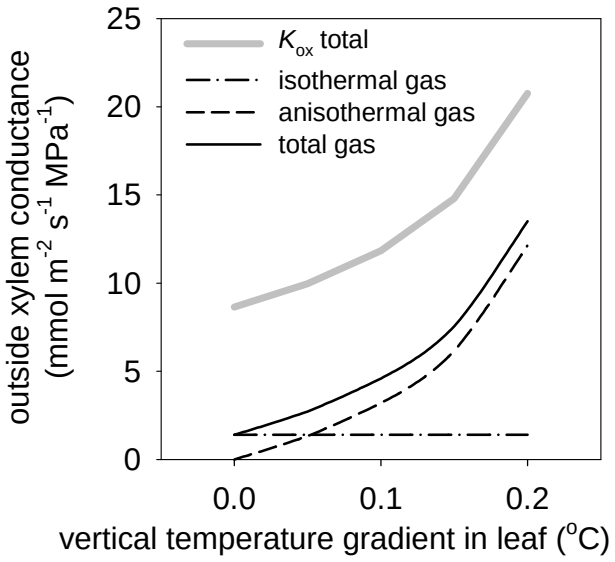


Figure 6. Effect of variation in vertical temperature gradients within leaves on outside-xylem hydraulic conductance, K_{ox} (thick grey line), and the components of K_{ox} that are attributable to gas phase transport that is independent of vertical gradients ("isothermal gas", dash-dot line), gas phase transport that is driven by vertical temperature gradients independent of water potential gradients ("anisothermal gas", dashed line) and the sum of those two gas phase components ("total gas", solid black line).

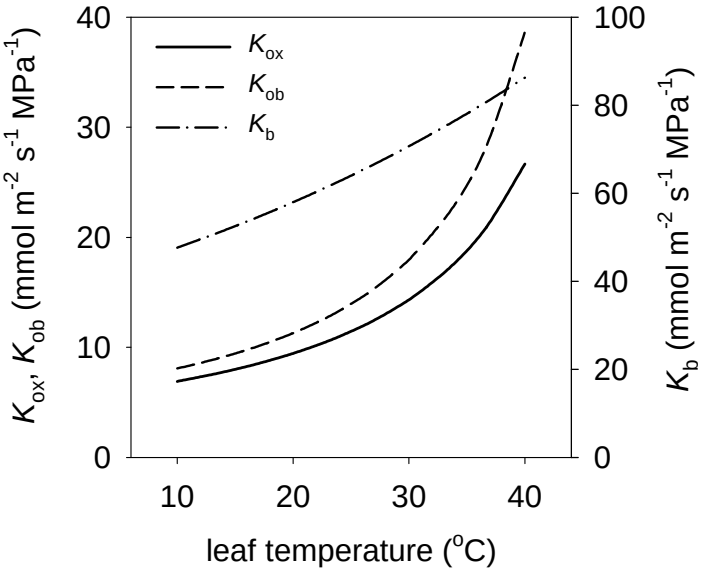


Figure 7. Effects of leaf temperature on outside-xylem hydraulic conductance, K_{ox}, and its components outside the bundle sheath (K_{ob}) and across the bundle sheath (K_b, right-hand axis). Note that K_b is plotted on a different scale than K_{ob} and K_{ox}.

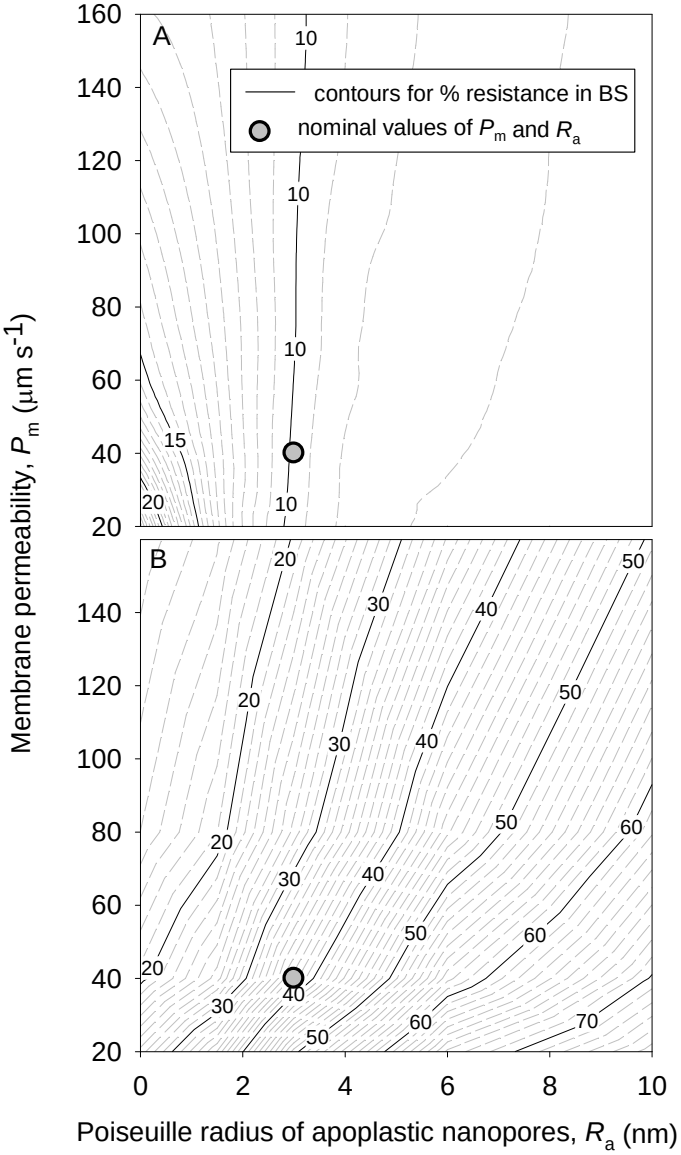


Figure 8. Effects of Poiseuille radius of apoplastic nanopores (R_a, horizontal axes) and osmotic water permeability of membranes (P_m, vertical axes) on percent of outside-xylem hydraulic resistance contributed by the bundle sheath (% resistance in BS, contours) for two sets of conditions: (a) default values for all parameters, as given in Tables III-IV, including no blockage of apoplastic transport in the bundle sheath; (b) default values for all parameters except that apoplastic pathways in the bundle sheath are blocked by a suberized layer or "Casparian strip". Grey symbols: location of default values for R_a and P_m (3 nm and 40 μm s^{-1}, respectively).

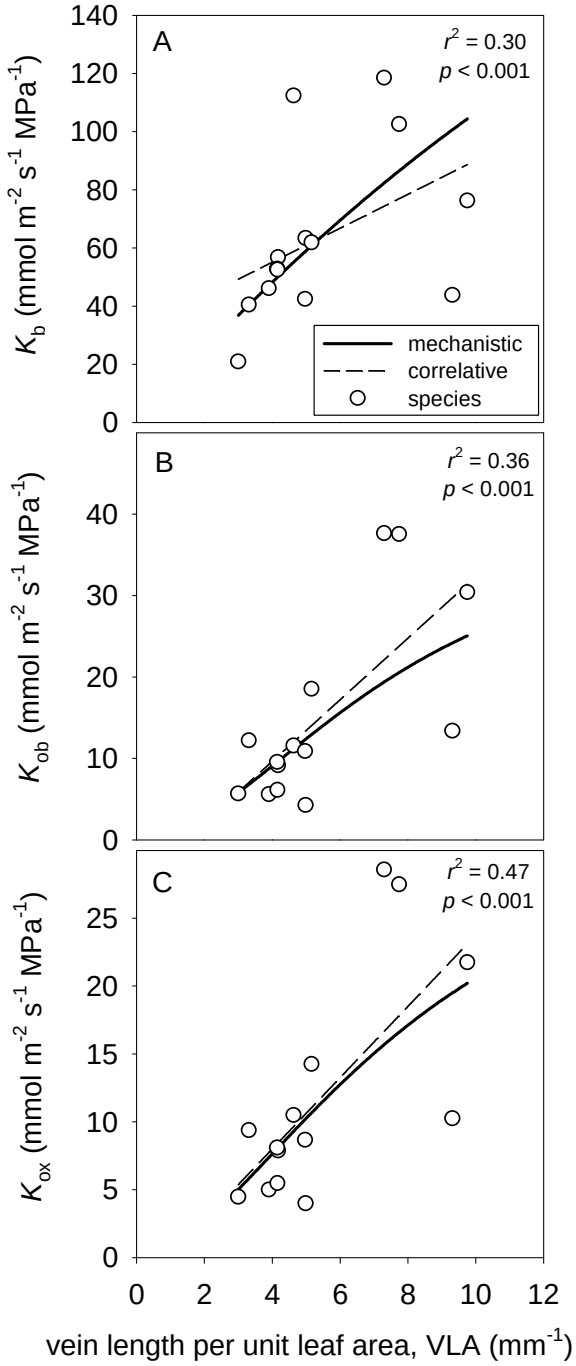


Figure 9. Effects of vein length per unit leaf area (VLA) on (a) hydraulic conductance across the bundle sheath, K_b; (b) outside-bundle sheath hydraulic conductance, K_{ob}, and (c) outside-xylem hydraulic conductance, K_{ox}. Solid lines are mechanistic relationships obtained by varying VLA in the model while holding all other parameters constant. Open symbols are values for each of 14 species, and dashed lines are the cross-species correlations between VLA and each conductance. r^2 and p values shown are for the cross-species correlations.

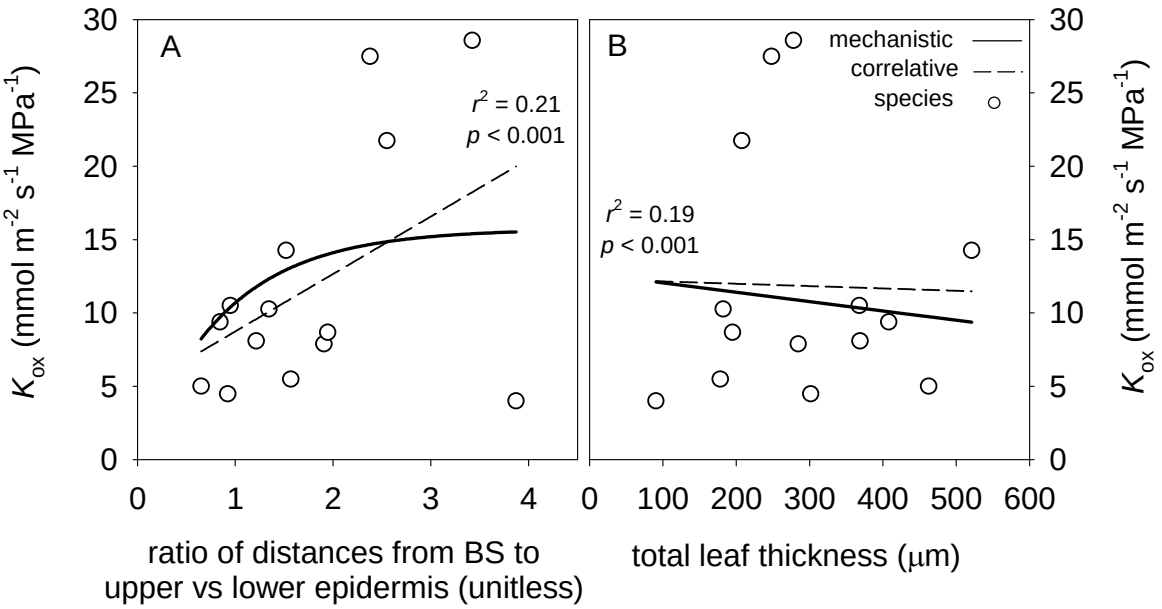


Figure 10. Effects of two anatomical parameters related to vertical distance between veins and the epidermis on outside-xylem hydraulic conductance, K_{ox}: A, the ratio of the distance between the bundle sheath [BS] and the upper epidermis to the distance to the lower epidermis, and B, total leaf thickness. Solid lines are mechanistic relationships obtained by varying the parameters shown on the x-axes in the model while holding all other parameters constant. Open symbols are values for each of 14 species, and dashed lines are the cross-species correlations between the parameters on the x-axes and each conductance. r^2 and p values shown are for the cross-species correlations.

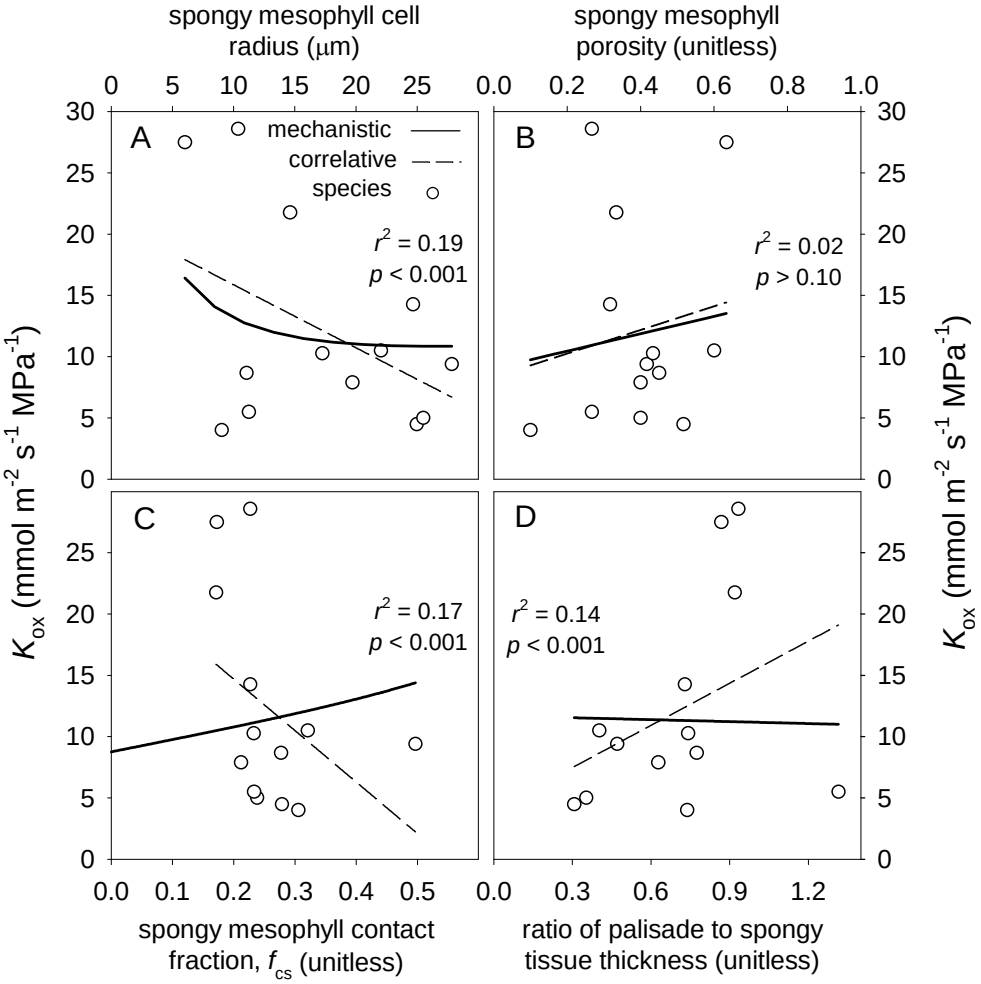


Figure 11. Effects of four anatomical parameters related to spongy mesophyll anatomy on outside-xylem hydraulic conductance, K_{ox}: A, spongy mesophyll cell radius, B, spongy mesophyll tissue airspace fraction, C, the fraction of spongy mesophyll surface area that is in contact with adjacent spongy cells, and D, the ratio of palisade tissue thickness to spongy tissue thickness. Solid lines are mechanistic relationships obtained by varying the parameters shown on the x-axes in the model while holding all other parameters constant. Open symbols are values for each of 14 species, and dashed lines are the cross-species correlations between the parameters on the x-axes and each conductance. r^2 and p values shown are for the cross-species correlations.

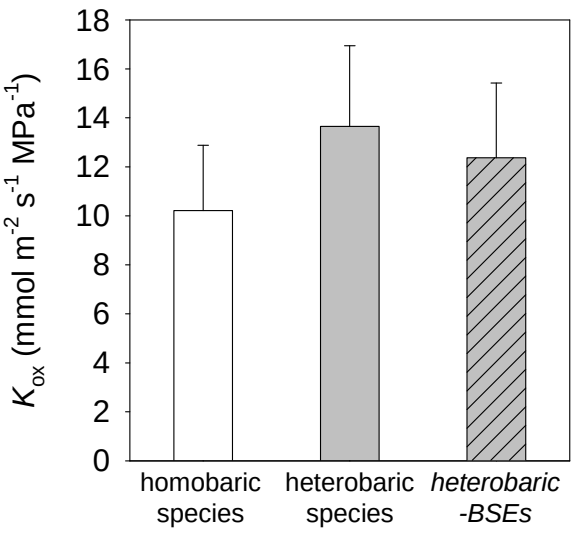


Figure 12. Differences in outside-xylem hydraulic conductance, K_{ox}, computed by the model for homobaric (white bar) and heterobaric species (grey bars). For heterobaric species, an additional simulation was performed in which nodes in the grid corresponding to bundle sheath extensions (BSEs) were assigned conductivities corresponding to the adjacent mesophyll (grey bar with hash marks; "heterobaric –BSEs").

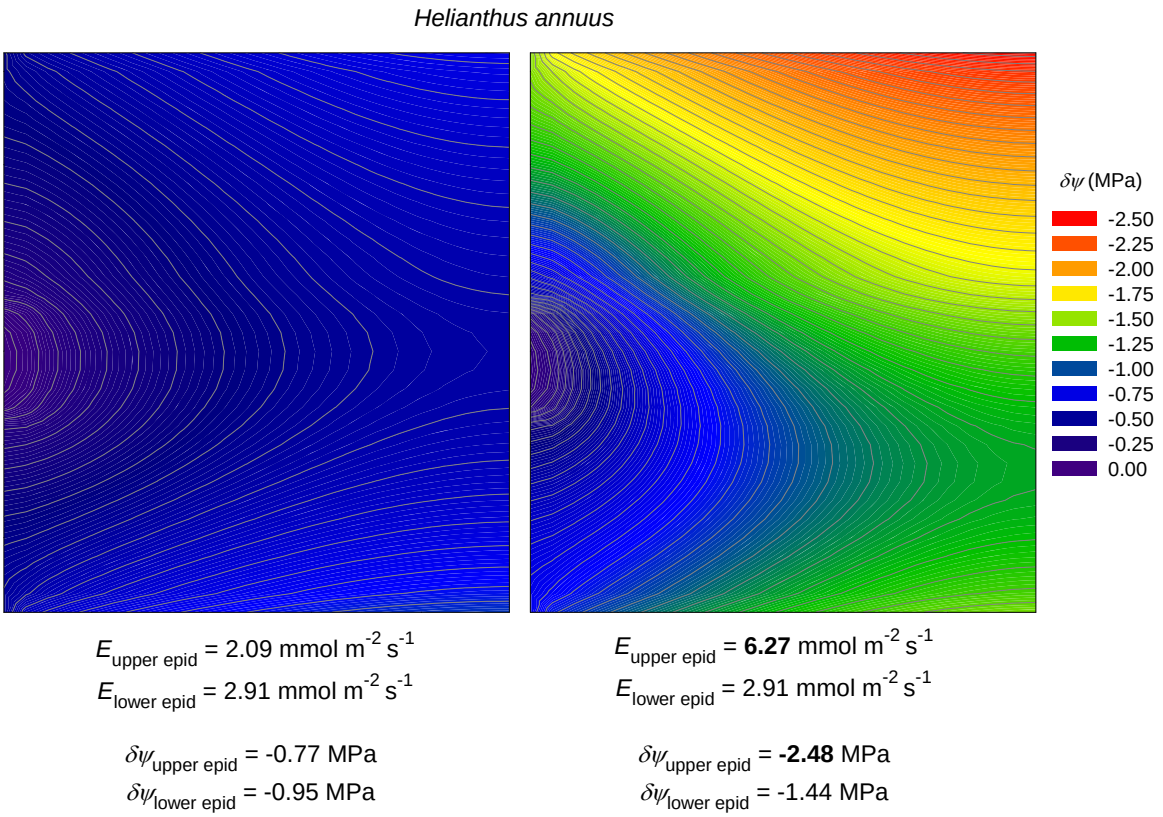
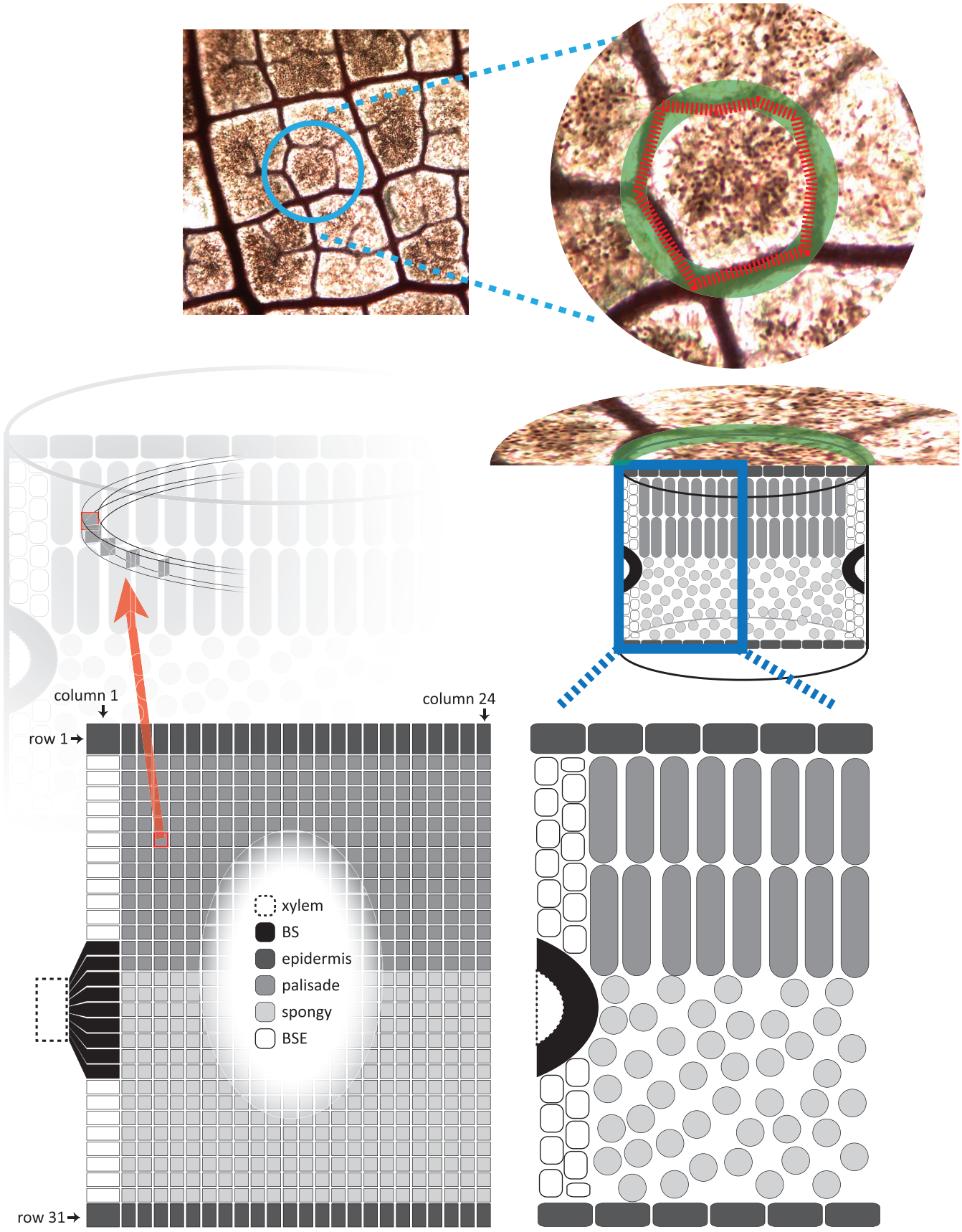


Figure 13. Simulation of the effect of increasing transpiration rate (*E*) three-fold at the upper surface of an amphistomatous leaf (*Helianthus annuus*) while holding it constant at the lower surface. Color contours represent water potential drawdowns from the xylem ($\delta\psi$); the areole margin (minor vein) is at left and the areole center is at right; the upper and lower surfaces are at top and bottom, respectively. Left panel: before the change in transpiration rate; right panel: after the change. Initial and final transpiration rates for both surfaces, and $\delta\psi$ values for the epidermal nodes at the center of the areole at both surfaces, are shown at bottom. These simulations used default values for all parameters as given in Tables III-IV.



Parsed Citations

Aasamaa K, Niinemets Ü, Sõber A (2005) Leaf hydraulic conductance in relation to anatomical and functional traits during Populus tremula leaf ontogeny. Tree Physiology 25: 1409-1418
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Barbour MM, Farquhar GD (2004) Do pathways of water movement and leaf anatomical dimensions allow development of gradients in H218O between veins and the sites of evaporation within leaves? Plant, Cell & Environment 27: 107-121
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Boyer JS (1985) Water transport. Annual Review of Plant Physiology 36: 473-516
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Brodribb TJ, Feild TS, Jordan GJ (2007) Leaf maximum photosynthetic rate and venation are linked by hydraulics. Plant Physiology 144: 1890-1898
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Brodribb TJ, Holbrook NM (2003) Stomatal closure during leaf dehydration, correlation with other leaf physiological traits. Plant Physiology 132: 2166-2173
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Brodribb TJ, Holbrook NM, Zwieniecki MA, Palma B (2005) Leaf hydraulic capacity in ferns, conifers and angiosperms: impacts on photosynthetic maxima. New Phytologist 165: 839-846
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Brodribb TJ, Jordan GJ (2008) Internal coordination between hydraulics and stomatal control in leaves. Plant, Cell & Environment 31: 1557-1564
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Brodribb TJ, Jordan GJ, Carpenter RJ (2013) Unified changes in cell size permit coordinated leaf evolution. New Phytologist 199: 559-570
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Buckley TN (2005) The control of stomata by water balance (Tansley Review). New Phytologist 168: 275-292
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Buckley TN (2015) The contributions of apoplastic, symplastic and gas phase pathways for water transport outside the bundle sheath in leaves. Plant, cell & environment 38: 7-22
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Buckley TN, Sack L, Gilbert ME (2011) The role of bundle sheath extensions and life form in stomatal responses to leaf water status. Plant Physiology 156: 962-973
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Carins Murphy MR, Jordan GJ, Brodribb TJ (2012) Differential leaf expansion can enable hydraulic acclimation to sun and shade. Plant, Cell & Environment 35: 1407-1418
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Carins Murphy MR, Jordan GJ, Brodribb TJ (2014) Acclimation to humidity modifies the link between leaf size and the density of veins and stomata. Plant, Cell & Environment 37: 124-131
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Chatelet DS, Wistrom CM, Purcell AH, Rost TL, Matthews MA (2011) Xylem structure of four grape varieties and 12 alternative hosts to the xylem-limited bacterium Xylella fastidiosa. Annals of Botany 108: 773-85

Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Cochard H, Nardini A, Coll L (2004) Hydraulic architecture of leaf blades: where is the main resistance? Plant, Cell & Environment 27: 1257-1267
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Cochard H, Venisse JS, Barigah TS, Brunel N, Herbette S, Guilliot A, Tyree MT, Sakr S (2007) Putative role of aquaporins in variable hydraulic conductance of leaves in response to light. Plant Physiology 143: 122-133
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Doelger J, Rademaker H, Liesche J, Schulz A, Bohr T (2014) Diffusion and bulk flow in phloem loading: A theoretical analysis of the polymer trap mechanism for sugar transport in plants. Physical Review E 90
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Eijkel JC, Van Den Berg A (2005) Nanofluidics: what is it and what can we expect from it? Microfluidics and Nanofluidics 1: 249-267
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Evans J, Caemmerer S, Setchell B, Hudson G (1994) The Relationship Between CO_2 Transfer Conductance and Leaf Anatomy in Transgenic Tobacco With a Reduced Content of Rubisco. Functional Plant Biology 21: 475-495
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Fahlén J, Salmén L (2004) Pore and Matrix Distribution in the Fiber Wall Revealed by Atomic Force Microscopy and Image Analysis. Biomacromolecules 6: 433-438
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Farquhar GD, Raschke K (1978) On the Resistance to Transpiration of the Sites of Evaporation within the Leaf. Plant Physiology 61: 1000-1005
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Feild TS, Brodribb TJ (2013) Hydraulic tuning of vein cell microstructure in the evolution of angiosperm venation networks. New Phytologist 199: 720-726
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Fleischer A, O'Neill MA, Ehwald R (1999) The Pore Size of Non-Graminaceous Plant Cell Walls Is Rapidly Decreased by Borate Ester Cross-Linking of the Pectic Polysaccharide Rhamnogalacturonan II. Plant Physiology 121: 829-838
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Flexas J, Scoffoni C, Gago J, Sack L (2013) Leaf mesophyll conductance and leaf hydraulic conductance: an introduction to their measurement and coordination. Journal of experimental botany 64: 3965-3981
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Guyot G, Scoffoni C, Sack L (2012) Combined impacts of irradiance and dehydration on leaf hydraulic conductance: insights into vulnerability and stomatal control. Plant, cell & environment 35: 857-871
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Hanba YT, Kogami H, Terashima I (2002) The effect of growth irradiance on leaf anatomy and photosynthesis in Acer species differing in light demand. Plant, Cell & Environment 25: 1021-1030
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

John GP, Scoffoni C, Sack L (2013) Allometry of cells and tissues within leaves. American journal of botany 100: 1936-1948
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Kennedy C, Šturcová A, Jarvis M, Wess T (2007) Hydration effects on spacing of primary wall cellulose microfibrils: a small angle X-

ray scattering study. Cellulose 14: 401-408
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Lersten NR (1997) Occurrence of endodermis with a casparian strip in stem and leaf. The Botanical Review 63: 265-272
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Lersten NR (1997) Occurrence of endodermis with a casparian strip in stem and leaf. Botanical Review 63: 265-272
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Martre P, Cochard H, Durand JL (2001) Hydraulic architecture and water flow in growing grass tillers (Festuca arundinacea Schreb.). Plant, Cell & Environment 24: 65-76
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

McCann MC, Wells B, Roberts K (1990) Direct visualization of cross-links in the primary plant-cell wall. J Cell Sci 96: 323-334
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Meidner H (1976) Water Vapour Loss from a Physical Model of a Substomatal Cavity. Journal of Experimental Botany 27: 691-694
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Meidner H (1983) Our Understanding of Plant Water Relations: The Bidder Lecture delivered at the Hull Meeting of the Society of Experimental Biology on 7 April 1983. Journal of experimental botany: 1606-1618
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Mertz RA, Brutnell TP (2014) Bundle sheath suberization in grass leaves: multiple barriers to characterization. Journal of Experimental Botany 65: 3371-3380
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Moghaddam PR, Wilman D (1998) Cell wall thickness and cell dimensions in plant parts of eight forage species. Journal of Agricultureal Science 131: 59-67
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Mott KA (2007) Leaf hydraulic conductivity and stomatal responses to humidity in amphistomatous leaves. Plant, Cell and Environment 30: 1444-1449
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Nardini A, Gortan E, Salleo S (2005) Hydraulic efficiency of the leaf venation system in sun-and shade-adapted species. Functional Plant Biology 32: 953-961
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Noblin X, Mahadevan L, Coomaraswamy I, Weitz DA, Holbrook NM, Zwieniecki MA (2008) Optimal vein density in artificial and real leaves. Proceedings of the National Academy of Sciences 105: 9140-9144
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Noblin X, Mahadevan L, Coomaraswamy IA, Weitz DA, Holbrook NM, Zwieniecki MA (2008) Optimal vein density in artificial and real leaves. Proceedings of the National Academy of Sciences of the United States of America 105: 9140-9144
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Pasquet-Kok J, Creese C, Sack L (2010) Turning over a new 'leaf': multiple functional significances of leaves versus phyllodes in Hawaiian Acacia koa. Plant, Cell & Environment 33: 2084-2100
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Rockwell F, Holbrook NM, Stroock AD (2014) The competition between liquid and vapor transport in transpiring leaves. Plant Physiology: doi: http://?dx.?doi.?org/?10.?1104/?pp.?114.?236323

Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sack L, Cowan PD, Jaikumar N, Holbrook NM (2003) The 'hydrology' of leaves: co-ordination of structure and function in temperate woody species. Plant, Cell and Environment 26: 1343-1356
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sack L, Frole K (2006) LEAF STRUCTURAL DIVERSITY IS RELATED TO HYDRAULIC CAPACITY IN TROPICAL RAIN FOREST TREES. Ecology 87: 483-491
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sack L, Holbrook NM (2006) Leaf hydraulics. Annual Review of Plant Biology 57: 361-381
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sack L, Melcher PJ, Zwieniechi MA, Holbrook NM (2002) The hydraulic conductance of the angiosperm leaf lamina: a comparison of three measurement methods. Journal of Experimental Botany 53: 2177-2184
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sack L, Scoffoni C (2013) Leaf venation: structure, function, development, evolution, ecology and applications in the past, present and future. New Phytologist 198: 983-1000
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sack L, Scoffoni C, John GP, Poorter H, Mason CM, Mendez-Alonzo R, Donovan LA (2013) How do leaf veins influence the worldwide leaf economic spectrum? Review and synthesis. Journal of Experimental Botany 64: 4053-4080
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sack L, Scoffoni C, John GP, Poorter H, Mason CM, Mendez-Alonzo R, Donovan LA (2014) Leaf mass per area is independent of vein length per area: avoiding pitfalls when modeling phenotypic integration. Journal of Experimental Botany in press
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sack L, Scoffoni C, Johnson DM, Buckley TN, Brodribb TJ (2015) The anatomical determinants of leaf hydraulic function. In U Hacke, ed, Functional and Ecological Xylem Anatomy. Springer International, Cham, Switzerland, pp 255-271
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sack L, Scoffoni C, McKown AD, Frole K, Rawls M, Havran JC, Tran H, Tran T (2012) Developmentally based scaling of leaf venation architecture explains global ecological patterns. Nature Communications 3: 837
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sack L, Streeter CM, Holbrook NM (2004) Hydraulic analysis of water flow through leaves of sugar maple and red oak. Plant Physiology 134: 1824-1833
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sack L, Tyree MT (2005) Leaf hydraulics and its implications in plant structure and function. Vascular transport in plants: 93-114
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Scafaro AP, Von Caemmerer S, Evans JR, Atwell BJ (2011) Temperature response of mesophyll conductance in cultivated and wild Oryza species with contrasting mesophyll cell wall thickness. Plant, Cell & Environment 34: 1999-2008
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Scoffoni C, Kunkle J, Pasquet-Kok J, Vuong C, Patel AJ, Montgomery RA, Givnish TJ, Sack L (2015) Light-induced plasticity in leaf hydraulics, venation, anatomy, and gas exchange in ecologically diverse Hawaiian lobeliads. New Phytologist
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Scoffoni C, McKown AD, Rawls M, Sack L (2012) Dynamics of leaf hydraulic conductance with water status: quantification and

analysis of species differences under steady state. Journal of Experimental Botany 63: 643-658
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Scoffoni C, Pou A, Aasamaa K, Sack L (2008) The rapid light response of leaf hydraulic conductance: new evidence from two experimental methods. Plant, Cell and Environment 31: 1803-1812
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Scoffoni C, Sack L (2014) Are leaves "freewheelin'"? Testing for a Wheeler-type effect in leaf xylem hydraulic decline. Plant, Cell & Environment in press
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Scoffoni C, Sack L (2015) Are leaves 'freewheelin'? Testing for a Wheeler-type effect in leaf xylem hydraulic decline. Plant, cell & environment 38: 534-543
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Simonin KA, Burns E, Choat B, Barbour MM, Dawson TE, Franks PJ (2014) Increasing leaf hydraulic conductance with transpiration rate minimizes the water potential drawdown from stem to leaf. Journal of Experimental Botany
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Slewinski TL, Anderson AA, Zhang C, Turgeon R (2012) Scarecrow plays a role in establishing Kranz anatomy in maize leaves. Plant and Cell Physiology 53: 2030-2037
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sommerville KE, Sack L, Ball MC (2012) Hydraulic conductance of Acacia phyllodes (foliage) is driven by primary nerve (vein) conductance and density. Plant, cell & environment 35: 158-168
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Sperry JS (2000) Hydraulic constraints on gas exchange. Agricultural and Forest Meteorology 104: 13-23
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Tyree MT, Cochard H (2003) Vessel contents of leaves after excision: a test of the Scholander assumption. Journal of Experimental Botany 54: 2133-2139
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Tyree MT, Yianoulis P (1980) The site of water evaporation from sub-stomatal cavities, liquid path resistances and hydroactive stomatal closure. Annals of Botany 45: 175-193
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Voicu MC, Cooke JE, Zwiazek JJ (2009) Aquaporin gene expression and apoplastic water flow in bur oak (Quercus macrocarpa) leaves in relation to the light response of leaf hydraulic conductance. Journal of Experimental Botany: erp239
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Wylie RB (1952) The Bundle Sheath Extension in Leaves of Dicotyledons. American Journal of Botany 39: 645-651
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Yang S, Tyree MT (1994) Hydraulic architecture of Acer saccharum and A. rubrum: comparison of branches to whole trees and the contribution of leaves to hydraulic resistance. Journal of Experimental Botany 45: 179-186
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Zsögön A, Alves Negrini AC, Peres LEP, Nguyen HT, Ball MC (2015) A mutation that eliminates bundle sheath extensions reduces leaf hydraulic conductance, stomatal conductance and assimilation rates in tomato (Solanum lycopersicum). New Phytologist 205: 618-626
Pubmed: Author and Title
CrossRef: Author and Title
Google Scholar: Author Only Title Only Author and Title

Supplemental File S1

Derivation of expressions for flow area, pathlength corrections, grid areas, grid pathlengths and volume averaging basis adjustment

To accompany "How does leaf anatomy influence water transport outside the xylem?" by TN Buckley et al.

Flow areas per unit bulk area, γ

For flow across membranes, the potential area for transport may be greater than the simple cross-sectional area, or bulk area. For example, the curved surfaces of mesophyll cells present a greater area of membrane for transport than the simple projected areas of those cells in the direction of flow. However, the effective area is also reduced in proportion to the connectivity of mesophyll cells (the fraction of total surface area that is in contact betweenadjacent cells; f_c), and the complement of the tissue porosity (p_p). The correction factors (γ) are the ratios of actual contacting surface area to projected cross sectional area. For vertical transmembrane flow in palisade tissue, this is

(S1) $$\gamma_{m,pv} = (1 - p_p) f_c \frac{\frac{1}{2} 4\pi r_p^2}{\pi r_p^2} = 2(1 - p_p) f_c$$

The numerator is one-half of the surface area of a sphere, which is also the surface area of the curved end of a capsule. The denominator is the projected cross-sectional area of the cell. An identical expression also arises for both vertical and horizontal transmembrane flow in spongy tissues:

(S2) $$\gamma_{m,sv} = \gamma_{m,sh} = 2(1 - p_s) f_c$$

The surface area of a capsule is $4 \cdot \pi \cdot r_p^2 + 2 \cdot \pi \cdot r_p (h_p - 2r_p)$ and the cross sectional area along the long axis is $\pi \cdot r_p^2 + 2r_p(h_p - 2r_p)$. Therefore, for horizontal transmembrane flow in palisade tissue,

(S3) $$\gamma_{m,ph} = (1-p_p) f_c \frac{\frac{1}{2}\left(4\pi r_p^2 + 2\pi r_p (h_p - 2r_p)\right)}{\pi r_p^2 + 2r_p (h_p - 2r_p)} = \frac{\pi (1-p_p) f_c h_p}{2h_p - (4-\pi) r_p}$$

All other cell types are modeled as rectangular boxes with zero tissue porosity, so their ratios of projected and actual transmembrane flow areas are simply unity.

For apoplastic flow paths, the total area available for flow is approximately the product of cell circumference, cell wall thickness and cell wall porosity, whereas the bulk area is the cell cross sectional area divided by the complement of tissue porosity. For vertical apoplastic flow in the palisade, this gives

(S4) $$\gamma_{a,pv} = \frac{t_{ap} \cdot 2\pi r_p \cdot p_a}{\pi r_p^2 / (1-p_p)} = \frac{2 p_a (1-p_p) t_{ap}}{r_p}$$

For horizontal apoplastic flow in the palisade, this gives

(S5) $$\gamma_{a,ph} = \frac{p_a (1-p_p) t_{ap} (2\pi r_p + 2h_p)}{\pi r_p^2 + 2r_p (h_p - 2r_p)}$$

For horizontal or vertical flow in the spongy mesophyll, the result is analogous to $\gamma_{a,pv}$:

(S6) $$\gamma_{a,sv} = \gamma_{a,sh} = \frac{2 p_a (1-p_s) t_{as}}{r_s}$$

We modeled epidermal cells as rectangular boxes with square bases of width w_e and height h_e. The area correction for vertical apoplastic flow into the epidermis is thus

(S7) $$\gamma_{a,ev} = \frac{p_a 4 w_e t_{ae}}{w_e^2} = \frac{4 p_a t_{ae}}{w_e}$$

For horizontal flow, the result is

(S8) $$\gamma_{a,eh} = \frac{p_a(2w_e + 2h_e)t_{ae}}{w_e h_e} = 2p_a t_{ae}\left(\frac{1}{h_e} + \frac{1}{w_e}\right)$$

We modeled bundle sheath and BSE cells as cubes with width w_b and w_x, respectively, which gives results analogous to $\gamma_{a,ev}$:

(S9) $$\gamma_{a,xv} = \gamma_{a,xh} = \frac{4p_a t_{ax}}{w_x}$$, and

(S10) $$\gamma_{a,b} = \frac{4p_a t_{ab}}{w_b}.$$

Finally, for gas flow, the area correction is simply the tissue porosity: $\gamma_{g,p} = p_p$ and $\gamma_{g,s} = p_s$ for palisade and spongy mesophyll, and zero for all other tissues.

Flow pathlengths per unit bulk pathlength, β

For apoplastic flow in spongy mesophyll and horizontal apoplastic flow in palisade mesophyll, the direct route across a cell is twice its radius, whereas the minimum apoplastic route is half of the cell circumference, or π times its radius, corrected for mesophyll connectivity (f_c). The terms for radius cancel out, giving

(S11) $$\beta_{a,sh} = \beta_{a,sv} = \beta_{a,ph} = \tfrac{1}{2}(1 - f_c)\pi$$

For vertical apoplastic flow in palisade mesophyll, the direct and apoplastic routes are both longer than for horizontal flow by the cell height h_p minus twice the radius, which gives

(S12) $$\beta_{a,pv} = \frac{(1-f_c)\pi r_p + (h_p - 2r_p)}{2r_p + (h_p - 2r_p)} = 1 + ((1-f_c)\pi - 2) r_p / h_p$$

The actual and direct flow paths are equivalent for all other tissues and modes of flow, giving β = 1.

Grid areas (a) and pathlengths (l)

The grid is a set of 744 tissue bands delineated by 25 planes parallel to the epidermis, and 31 concentric cylinders centered on a vertical axis located at the center of the areole. The upper- and lower-most planes coincide with the upper and lower leaf surfaces, respectively, thus defining 24 "rows" of tissue bands (indicated below with subscripted indices *i*). The outermost cylinder coindicides with the lateral midpoint of the nearest minor vein, thus defining 31 "columns" of tissue bands (indicated below with indices *j*).

With three exceptions, the vertical and radial thicknesses of these tissue bands are identical to one another. Two of these exceptions are the uppermost and lowermost rows (i = 1 and 31, respectively), whose thicknesses are defined by the measured upper and lower epidermis thicknesses, respectively. All other rows are defined as 1/29th of the remaining leaf thickness (equal to the sum of measured palisade and spongy mesophyll tissue thicknesses, t_p and t_s). The third exception, which applies only in heterobaric species (those possessing bundle sheath extensions) is the outermost column (j = 1), whose thickness is defined as one-half of the measured bundle sheath extension width. In these species, the widths of all other tissue columns are equal to 1/23rd of the difference between areole radius (r_{areole}) and BSE half-width ($w_{x,tot}/2$). In homobaric species (which lack bundle sheath extensions), all columns have the same width, which is 1/24th of the areole radius. These three exceptions ensure that the volumes, horizontal areas and flow pathlengths involving the epidermis and/or BSEs are appropriate to the actual tissue dimensions. (Further corrections are required to accommodate the modeled geometry of the bundle sheath; these are described in the next section.)

The area for horizontal flow between tissue bands j and j+1 with thickness t_i ($t_i = t_e$ for epidermal rows or $(t_p + t_s)/29$ otherwise) is equal to the product of t_i and the circumference of the outer cylinder bounding the band at j+1. For heterobaric species, this area is

(S13) $$a_{h,i}[j; j+1] = t_i \cdot 2\pi\left(r_{areole} - \tfrac{1}{2} w_{x,tot}\right)\left(1 - (j-1)/23\right)$$

and for homobaric species, this area is

(S14) $$a_{h,i}[j; j+1] = t_i \cdot 2\pi r_{areole}\left(1 - j/24\right)$$

The area for vertical flow between bands at column j and rows i and i+1 is equal to the vertical projected area of the band at column j. This equals the difference between the areas of circles defined by the outer and inner radial boundaries of column j. For the outermost column (j = 1) in heterobaric species, this area is

(S15) $$a_{v,j=1} = \pi r_{areole}^2 - \pi\left(r_{areole} - \tfrac{1}{2} w_{x,tot}\right)^2$$

For all other columns in heterobaric species, the area is

(S16) $$\begin{aligned} a_{v,j>1} &= \pi\left[\left(r_{areole} - \tfrac{1}{2} w_{x,tot}\right)\left(1 - (j-1)/23\right)\right]^2 - \pi\left[\left(r_{areole} - \tfrac{1}{2} w_{x,tot}\right)\left(1 - j/23\right)\right]^2 \\ &= \pi\left(r_{areole} - \tfrac{1}{2} w_{x,tot}\right)^2 (47 - 2j)/529 \end{aligned}$$

For all columns in homobaric species, the area is

(S17) $$a_{v,j} = \pi r_{areole}^2 (49 - 2j)/576$$

The direct flow pathlengths between adjacent bands are computed as the distances between the vertical and radial midpoints of those bands. Thus, the direct pathlength between the upper epidermis (i = 1) and the row of tissue bands directly below it (i = 2) equals one-half of the upper

epidermis thickness (t_{eu}) plus one-half of the thickness of one non-epidermis band, which as described above is 1/29th of the sum of palisade and spongy tissue thicknesses. This flow path is

(S18) $$l_v[i=1; i=2] = \tfrac{1}{2} t_{eu} + \tfrac{1}{29}\left(t_p + t_s\right)$$

Similarly, the direct flow pathlength between rows 30 and 31 (the lower epidermis) is

(S19) $$l_v[i=30; i=31] = \tfrac{1}{2} t_{el} + \tfrac{1}{29}\left(t_p + t_s\right)$$

The direct vertical flow pathlengths between all other rows is simply

(S20) $$l_{v,2 \le i \le 30} = \tfrac{1}{29}\left(t_p + t_s\right)$$

The direct horizontal flow pathlength between the outermost tissue band column (j = 1) and the adjacent column (j = 2) differs for heterobaric and homobaric species. For heterobaric species, the value is one-half of the bundle sheath extension half-width plus one-half of 1/23 of the remainder of the areole radius:

(S21) $$l_h[j=1; j=2] = \tfrac{1}{4} w_{x,tot} + \tfrac{1}{46}\left(r_{areole} - \tfrac{1}{2} w_{x,tot}\right)$$

For connections between all other adjacent columns in heterobaric species, the value is

(S22) $$l_{h,\, j \ge 2} = \tfrac{1}{23}\left(r_{areole} - \tfrac{1}{2} w_{x,tot}\right)$$

For homobaric species, the direct horizontal flow pathlength between any two adjacent columns is simply 1/24th of the areole radius:

(S23) $$l_h = r_{areole}/24$$

Estimating the number of grid rows for each tissue type

We estimated the number of grid rows for different tissue types based on measured tissue thicknesses, as follows. The number of bundle sheath rows (n_{bs}) was the greater of 1 and the quantity $29 \cdot h_{bs}/(t_p + t_s)$, rounded to the nearest whole number. This recognises that the palisade and spongy mesophyll tissue thicknesses combined ($t_p + t_s$) occupy 29 grid rows in total. The number of rows between the BS and the upper epidermis, n_{xu}, was computed as $(29 - n_{bs}) \cdot h_{xu}/(h_{xu} + h_{xl})$ (rounded to the nearest whole number), where h_{xu} and h_{xl} are the distances from the bundle sheath to the upper and lower epidermis, respectively; the number of rows between the BS and the lower epidermis, n_{xl}, was then $29 - n_{bs} - n_{xu}$. The number of palisade rows (n_p) was $29 \cdot t_p/(t_p + t_s)$, rounded to the nearest whole number, and the numer of spongy mesophyll rows (n_s) was $29 - n_p$.

Corrections to account for bundle sheath geometry

We modeled the bundle sheath as the space between the radial faces of two torus-like objects: one is actually an elliptic torus, and the other is a similar solid of revolution that is nested within the elliptic torus but has dimensions such that the distance from its surface to that of the elliptic torus is everywhere identical. The elliptic torus represents the outer face of the BS (the face farther from the xylem), the smaller (inner) torus-like object is the inner face, and the constant distance between the two faces represents the constant thickness of the BS itself. The outer face contacts mesophyll and BSE tissues. However, because the BS is represented in the grid as simply a stack of cylindrical tissue bands in the outermost column of tissue bands, the total area of the BS is not accurately represented by the grid areas computed as described in the preceding section. We therefore corrected the values for BS bulk conductivity applied to the grid in such a way that the total hydraulic conductance out of the BS accurately reflects the toroidal model described above. In this section, we describe how the relevant areas and corrections were calculated.

The area of the inner and outer faces of the BS can be computed using Pappus' Centroid Theorem, which states that the surface area of a surface of revolution created by revolving a

curve about an axis is equal to the product of the arc length of the curve and the distance travelled during the revolution by the curve's centroid (the point coinciding with the geometric average of all points in the curve). To compute these values, we therefore require the appropriate arc lengths, centroids and radii of revolution. The major radius of both tori is equal to the areole radius. The vertical radius of the ellipse (the "tube") for the outer torus is one-half of the measured height of the bundle sheath (h_{bs}), and the horizontal radius of that ellipse is computed from h_{bs} and the bundle sheath perimeter (p_{bs}) as $p_{bs}/\pi - h_{bs}/2$. The two radii of the ellipse for the inner torus are smaller than the analogous values for the outer torus by an amount equal to the measured bundle sheath cell thickness (t_{bs}).

The arc length for the outer face is thus simply $p_{bs}/2$, and the arc length for the inner face is $\pi((p_{bs}/\pi - h_{bs}/2 - t_{bs}) + (h_{bs}/2 - t_{bs}))/2 = \pi(p_{bs}/\pi - 2\cdot t_{bs})/2 = p_{bs}/2 - \pi\cdot t_{bs}$. It is easily shown that the centroid for the outer face is located at a distance $4(p_{bs}/\pi - h_{bs}/2)/(3\pi)$ from the edge of the areole, and the centroid for the inner face is located at a distance $4(p_{bs}/\pi - h_{bs}/2 - t_{bs}))/(3\pi)$. The distance travelled by these centroids during revolution is 2π times the difference between r_{areole} and each of these values. Thus, the area of the outer face is

(S24) $$a_{bs,out} = \left[\frac{p_{bs}}{2}\right]\cdot\left[2\pi\left(r_{areole} - \frac{4}{3\pi}\left(\frac{p_{bs}}{\pi} - \frac{h_{bs}}{2}\right)\right)\right]$$

and the area of the inner face is

(S25) $$a_{bs,in} = \left[\frac{p_{bs}}{2} - \pi t_{bs}\right]\cdot\left[2\pi\left(r_{areole} - \frac{4}{3\pi}\left(\frac{p_{bs}}{\pi} - \frac{h_{bs}}{2} - t_{bs}\right)\right)\right]$$

The total computed grid area for contact between the BS and mesophyll in heterobaric species is n_{bs} times the horizontal projected area for contact between the outermost column and the adjacent column, which from Eqn S13 is

(S26) $$a_{grid,bs-mes} = n_{bs} \cdot \frac{(t_p + t_s)}{29} \cdot 2\pi\left(r_{areole} - \tfrac{1}{2} w_{x,tot}\right)$$

For homobaric species, this area (from Eqn S14) is

(S27) $$a_{grid,bs-mes} = n_{bs} \cdot \frac{(t_p + t_s)}{29} \cdot 2\pi r_{areole}\left(\frac{23}{24}\right)$$

The total computed grid area for BS to BSE contact in heterobaric species is equal to twice the vertical projected area of a tissue band in the outermost column, which from Eqn S15 is

(S28) $$a_{grid,bs-bse} = 2\left(\pi r_{areole}^2 - \pi\left(r_{areole} - \tfrac{1}{2} w_{x,tot}\right)^2\right)$$

For homobaric species, from Eqn S17 this area is

(S29) $$a_{grid,bs-mes,vert} = 2\pi r_{areole}^2 \, 47/576$$

We assumed that the fraction of the total outer BS surface area in contact with the BSEs was equal to the BSE width divided by one-half of the BS perimeter ($2 \cdot w_{x,tot}/p_{bs}$), so that the total BS-BSE surface area was $2 \cdot w_{x,tot} \cdot a_{bs,out}/p_{bs}$. Thus, when calculating conductances for vertical transport between BS and BSE nodes in heterobaric species, the bulk flow area computed from Eqn x was corrected by the ratio

(S30) $$\frac{(2w_{x,tot} / p_{bs}) a_{bs,out}}{a_{grid,bs-bse}}$$

For conductances for horizontal transport between BS and mesophyll nodes in heterobaric species, the bulk flow area was corrected by the following ratio:

(S31) $$\frac{\left(1-2w_{x,tot}/p_{bs}\right)a_{bs,out}}{a_{grid,bs-bse}}$$

The analogous corrections for homobaric species are identical except that the BSE width $w_{x,tot}$ is replaced by twice the width of the outermost column of tissue bands in these species, or $2 \cdot r_{areole}/24$:

(S32) $$\frac{\left(r_{areole}/6p_{bs}\right)a_{bs,out}}{a_{grid,bs-bse}}$$, and

(S33) $$\frac{\left(1-r_{areole}/6p_{bs}\right)a_{bs,out}}{a_{grid,bs-bse}}.$$

Computing volumes for each node

To compute volumes for each node, we consider each node to represent a three-dimensional annulus within the areole, bounded horizontally by cylinders and vertically by planes. For mesophyll nodes, the cylinders are chosen to bisect the lines connecting each adjacent node horizontally, and the planes are chosen to bisect the lines connecting each adjacent node vertically. For the upper and lower rows of nodes, which represent epidermis, we used measured upper and lower epidermis thicknesses (t_{eu} and t_{el}) to compute volumes. For BSE nodes, we used the measured BSE half-width ($w_{x,tot}/2$) to compute volumes. The calculations of hydraulic conductances presented in this study did not require computation of bundle sheath node volumes.

Example of spurious differences in BS and outside-BS hydraulic conductances arising from defining outside-xylem hydraulic conductance in terms of the average water potential of the entire symplast

In experimental studies, K_{ox} is defined in terms of leaf water potential, which equals the volume-weighted average water potential of the entire outside-xylem compartment (ψ_{ox}), provided that negligible water leaves the xylem during equilibration after excision. Thus,

(S34) $K_{ox} = E/|\psi_{ox}|$.

In this study, we sought to partition K_{ox} into two serial pathways: the BS (with conductance K_b) and the outside-BS compartment (with conductance K_{ob}), such that

(S35) $$\frac{1}{K_{ox}} = \frac{1}{K_b} + \frac{1}{K_{ob}}.$$

where K_b is calculated from the water potential drawdown across the BS, or $\delta\psi_{bn}$ (the drawdown to grid nodes immediately adjacent to, or neighboring, BS nodes; hence the subscript *bn*):

(S36) $K_b = E/\delta\psi_{bn}$.

Combining S34-S36 gives K_{ob} as

(S37) $K_{ob} = E/(|\psi_{ox}| - \delta\psi_{bn})$.

The value of K_{ob} given by Eqn S37 is not uniquely defined by outside-BS water transport properties, however. Imagine two leaves that are identical in every respect except for the value of K_b; specifically, suppose leaf A has K_b twice as large as leaf B. The computed values of K_{ob} will differ for these two leaves, as demonstrated in the example shown in the table below, and the difference will depend on the volume fractions of the BS and outside-BS compartments. This spurious difference in calculated K_{ob} can be traced to the fact that K_{ob} is defined in terms of a water potential (ψ_{ox}) that includes some tissues (the BS) upstream of the tissues whose transport properties are meant to be characterised by K_{ob}. In the example below, this leads to a 63% difference in K_{ob} between the two leaves, even though they have identical outside-BS transport properties.

parameter	leaf A	leaf B
fraction of OX volume that is in BS (f_b)	0.1	
fraction of OX volume that is distal to BS (f_{ob})	0.9	
transpiration rate (E)	10	
water potential drawdown from outer edge of BS to site of average water potential of outside-BS tissues ($\delta\psi_{ob}$)	0.4	
BS hydraulic conductance (K_b)	50	25
water potential at outer edge of BS (ψ_{bn})	$-E/K_b = 0.2$	0.4
water potential of BS (ψ_b)	$\psi_{bn}/2 = -0.1$	-0.2
average water potential of outside-BS tissues (ψ_{ob})	$\psi_{bn} - \delta\psi_{ob} = -0.5$	-0.6
outside-xylem water potential (ψ_{ox})	$f_b \cdot \psi_b + f_{ob} \cdot \psi_{ob} = -0.46$	-0.56
outside-xylem hydraulic conductance (K_{ox})	$E/\lvert\psi_{ox}\rvert = 21.74$	17.86
outside-BS hydraulic conductance (K_{ob})	$1/(1/K_{ox}-1/K_b) = 38.46$	62.54
% spurious difference in K_{ob}	63%	

We addressed this issue by defining K_{ox} in terms of the volume-weighted water potential drawdown to all tissues outside the BS ($\delta\psi_{ob}$; Eqn 10 in the main text).

Table S1. Flow areas per unit bulk area (γ) used to calculate bulk conductivities from intrinsic conductivities.

tissue	flow direction	symbol(s)	formula
apoplastic flow			
palisade mesophyll	vertical	$\gamma_{a,pv}$	$2p_a(1-p_p)t_{ap}/r_p$
palisade mesophyll	horizontal	$\gamma_{a,ph}$	$\frac{p_a(1-p_p)t_{ap}(2\pi r_p+2h_p)}{\pi r_p^2+2r_p(h_p-2r_p)}$
spongy mesophyll	both	$\gamma_{a,s}$	$2p_a(1-p_s)t_{as}/r_s$
epidermis	vertical	$\gamma_{a,ev}$	$4p_a t_{ae}/w_e$
epidermis	horizontal	$\gamma_{a,eh}$	$2p_a t_{ae}(h_e^{-1}+w_e^{-1})$
bundle sheath extensions	both	$\gamma_{a,x}$	$4p_a t_{ax}/w_x$
bundle sheath	-	$\gamma_{a,b}$	$4p_a t_{ab}/w_b$
transmembrane flow			
palisade mesophyll	vertical	$\gamma_{m,pv}$	$2(1-p_p)f_c$
palisade mesophyll	horizontal	$\gamma_{m,ph}$	$\frac{\pi(1-p_p)f_c h_p}{2h_p-(4-\pi)r_p}$
spongy mesophyll	both	$\gamma_{m,s}$	$2(1-p_s)f_c$
other	both	γ_m	1
gas phase flow			
palisade mesophyll	both	$\gamma_{g,p}$	p_p
spongy mesophyll	both	$\gamma_{g,s}$	p_s
other	both	γ_g	1

Table S2. Flow pathlengths per unit direct pathlength (β) used to calculate bulk conductivities from intrinsic conductivities.

tissue	flow direction	symbol(s)	formula
apoplastic flow			
palisade mesophyll	vertical	$\gamma_{\mathrm{a,pv}}$	$1+\left(\left(1-f_c\right)\pi-2\right)r_p/h_p$
palisade mesophyll	horizontal	$\gamma_{\mathrm{a,ph}}$	$\frac{1}{2}\left(1-f_c\right)\pi$
spongy mesophyll	both	$\gamma_{\mathrm{a,s}}$	$\frac{1}{2}\left(1-f_c\right)\pi$
other	any	γ_{a}	1
transmembrane flow			
all	any	γ_{m}	1
gas phase flow			
all	any	γ_{g}	1